

Airway Corynebacterium interfere with Streptococcus pneumoniae and Staphylococcus aureus infection

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Introduction

- Streptococcus pneumoniae colonizes the nasopharynx of 5-70% of the population worldwide and is an important precursor of lower airway infection.
- *S. pneumoniae* is a leading infectious cause of death in children under 5 years of age and is the most common cause of community-acquired bacterial pneumonia.
- Current pneumococcal vaccines target up to 23 serotypes of *S. pneumoniae*, however, with over 100 serotypes in circulation and only 60-70% effectives in covered serotypes, this provides only partial protection.
- Staphylococcus aureus asymptomatically colonizes the anterior nares of 20-30% of the population and is associated with an increased risk of distant infection including skin and soft tissue infections, endocarditis, bacteremia, and pneumonia.
- There is currently no vaccine for *S. aureus* and prevention strategies are limited to hygiene and contact prevention.
- Corynebacterium are a commensal bacterium in the airway which are correlated with reduced S. aureus and S. pneumoniae colonization and promotion of a more stable airway microbiome.
- Here we investigated the potential of *Corynebacterium* colonization as a preventive strategy against pathogen infection.

C. pseudodiphtheriticum protects against airway pathogen infection in mice

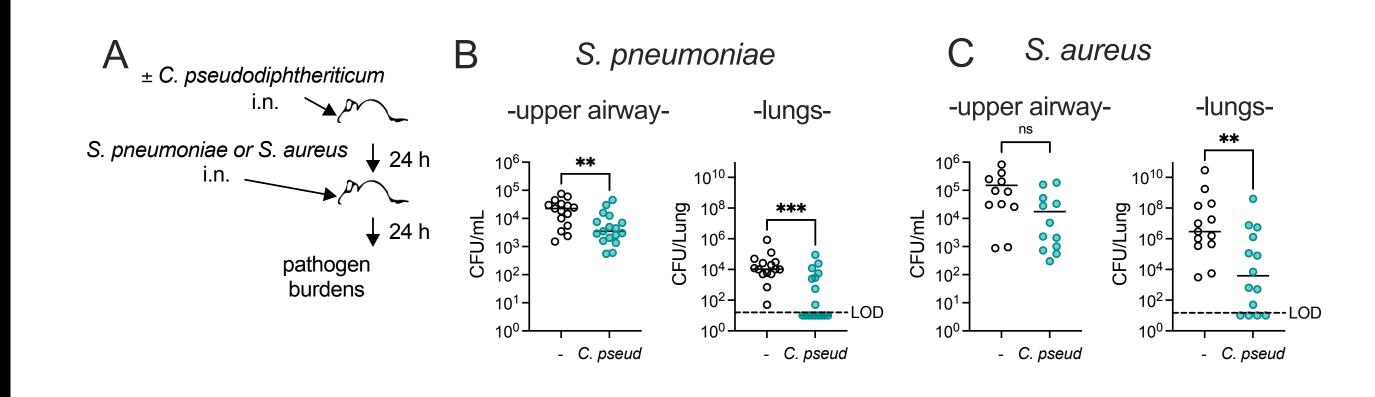


Figure 1. Exposure to *C. pseudodiphtheriticum* reduces *S. pneumoniae* and *S. aureus* respiratory tract

(A) Pathogen burdens detected in mice at 24 hours post infection with or without pre-exposure to *C. pseudodiphtheriticum* (*C. pseud*) i.n. Mice were treated with antibiotics for two weeks prior to bacterial exposures. **p<.01, ***p<.001, Mann-Whitney *U* test. Data are pooled from three independent experiments. LOD indicates the limit of detection for CFUs.

Corynebacterium secreted lipase selectively inhibits S. pneumoniae growth

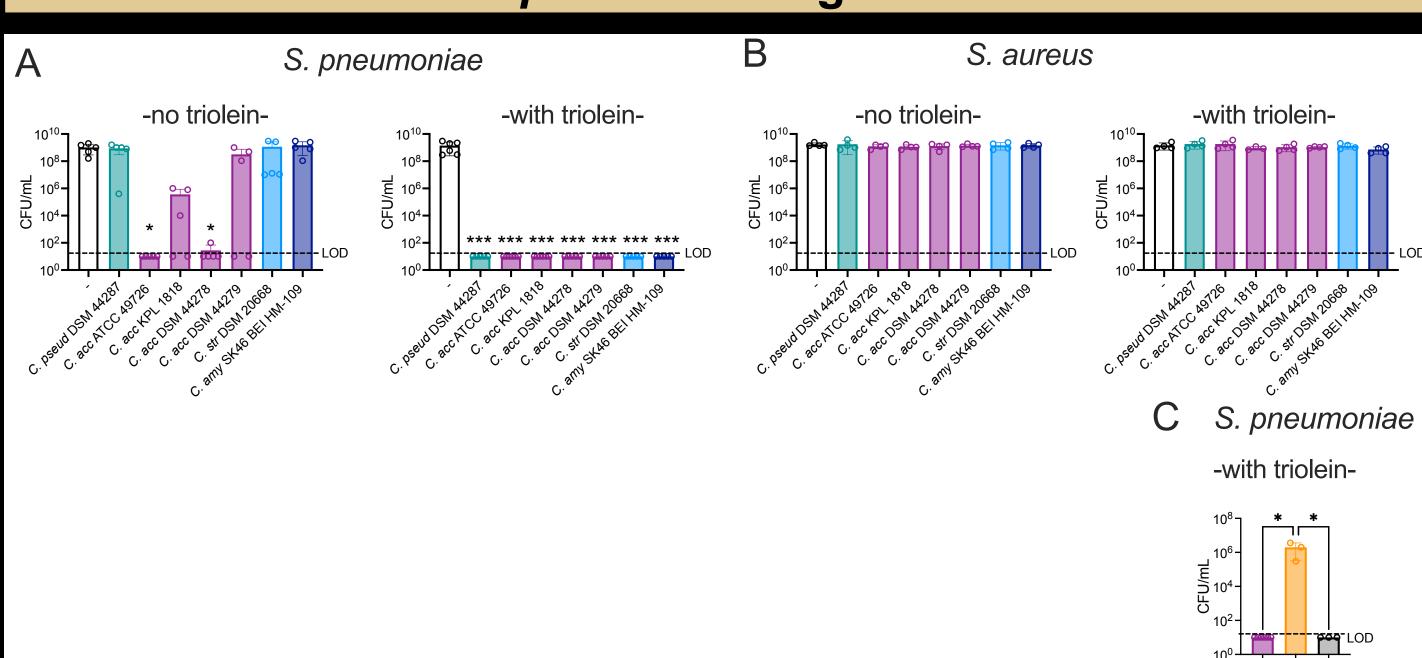


Figure 2. Corynebacterium secreted lipase inhibits *S. pneumoniae* growth without affecting *S. aureus*.

(A) Growth of *S. pneumoniae* following pre-spreading plates with supernatants from *Corynebacterium* strains as indicated grown with 1% Tween 80 alone (no triolein) or supplemented with 180 mg/mL triolein (with triolein). (B) Growth of *S. aureus* as in (A). (C) Growth of *S. pneumoniae* as for (A) following pre-spreading with supernatants from *C. accolens* WT (*C. acc*), a *C. accolens* mutant deficient in *lipS1* (*C. acc*^{ΔlipS1}), or a *lipS1* complemented strain (*C. acc*^{compl}), *p<.05, ***p<.001, Kruskal-Wallis test with Dunn's post hoc analysis. Data are pooled from 3-4 independent experiments. LOD indicates the limit of detection.

C. pseudodiphtheriticum reduces pathogen adherence to human respiratory tract epithelial cells

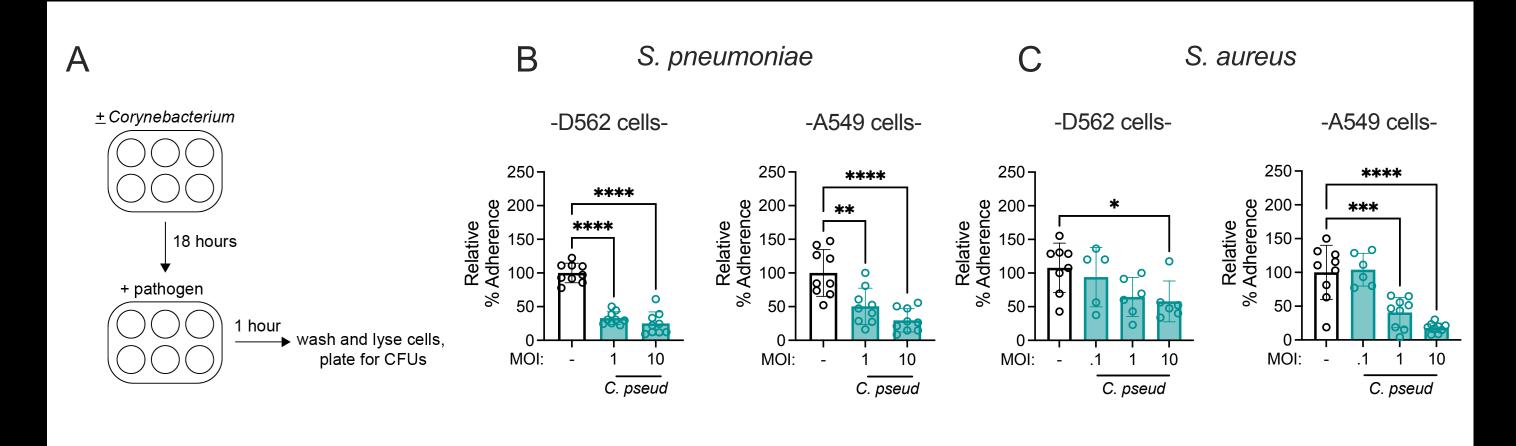


Figure 3. *C. pseudodiphtheriticum* colonization reduces adherence of *S. pneumoniae* and *S. aureus* to human respiratory tract epithelial cells. (A) Cell adherence assay schematic. (B) Percent adherence of *S. pneumoniae* on D526 and A549 cells at 1 hour post-infection with or without pre-colonization of epithelial cells with *C. pseudodiphtheriticum* for 18 hours at the indicated MOI. (D) Percent adherence of *S. aureus* as for (C). *p<.05, **p<.01, ****p<.0001, one-way ANOVA with Dunnett's post hoc analysis. Data are pooled from two independent experiments (A) or three independent experiments (C-E) with 3 replicates per condition.

C. accolens reduces pathogen adherence to epithelial cells in a lipase-independent manner

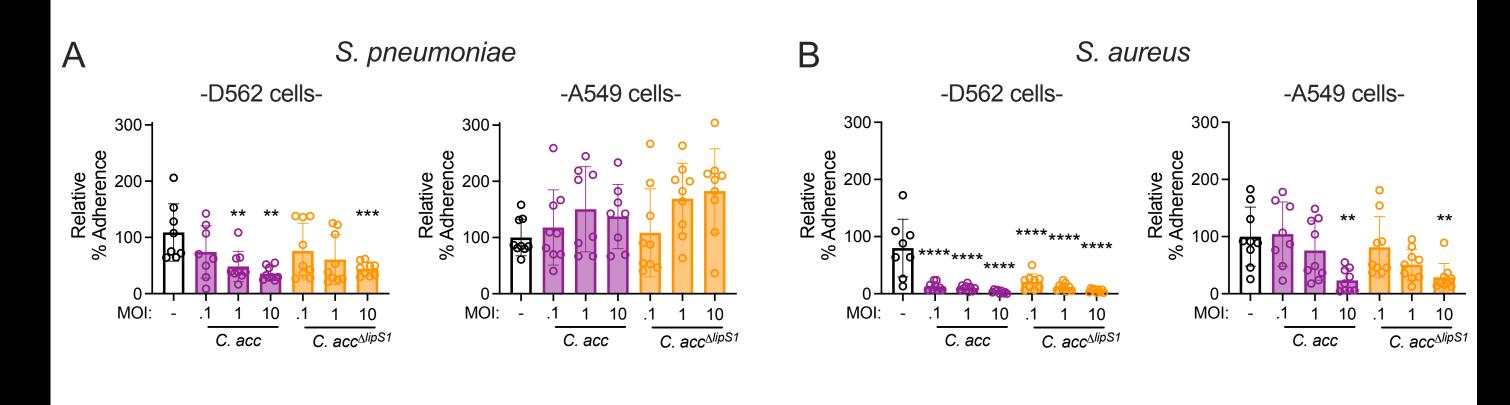


Figure 4. *C. accolens* colonization reduces pathogen adherence to human respiratory tract epithelial cells in a lipase-independent manner. (A) Percent adherence of *S. pneumoniae* on D526 and A549 cells at 1 hour post-infection with or without pre-colonization of epithelial cells with *C. accolens* WT (*C. acc*) or *lipS1* deficient *C. accolens* (*C. acc^{\Delta lipS1}*) for 18 hours at the indicated MOI. (B) Percent adherence of *S. aureus* as for (A). (**p<.01, ***p<.001, one-way ANOVA with Dunnett's post hoc analysis. Data are pooled from two independent experiments (A) or three independent experiments (B-D) with 3 replicates per condition.

Epithelial cell colonization inhibition requires live Corynebacterium

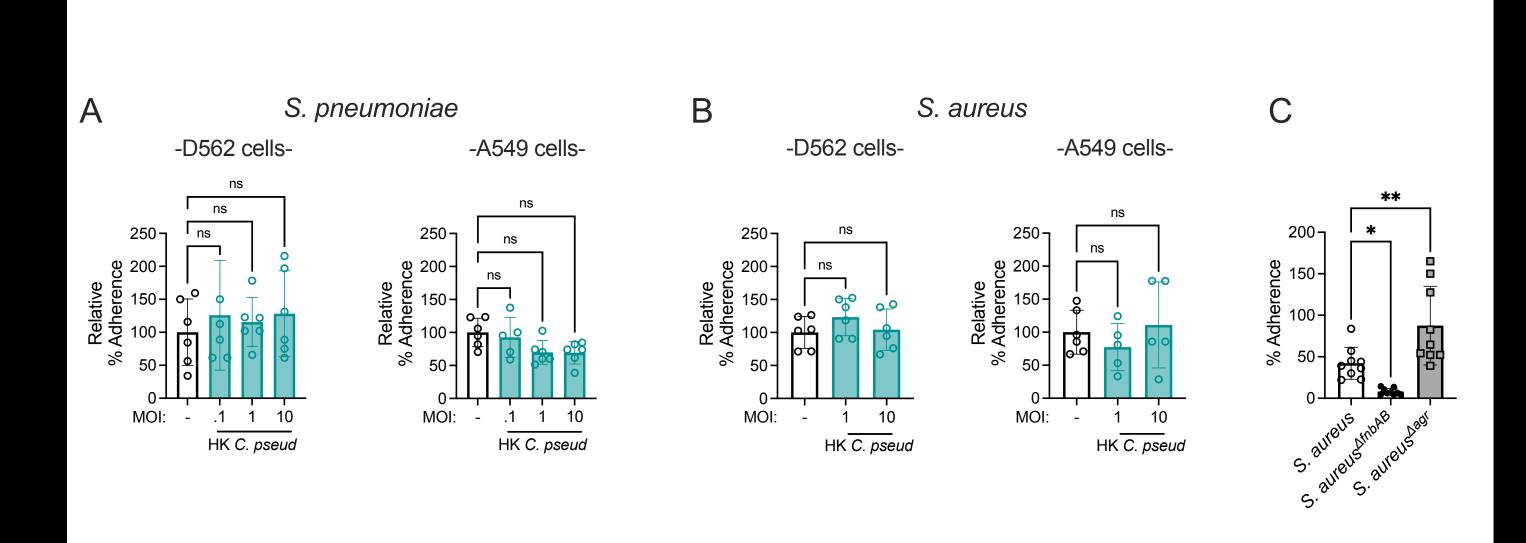


Figure 5. Corynebacterium colonization interference requires live bacteria and is sensitive to *S. aureus* adherence capacity. (A) Percent adherence of *S. pneumoniae* on D526 and A549 cells 1 hour post-infection with or without 18-hour pre-exposure to heat-killed *C. pseudodiphtheriticum* (HK *C. pseud*) at the indicated MOI. (B) Percent adherence of *S. aureus* MRSA strain USA300 as for (A). (C) Percent adherence of WT *S. aureus*, *fnbAB* deficient *S. aureus*, and *agr* deficient *S. aureus* to A549 cells 1 hour post-infection. *p<.05, **p<.01, one-way ANOVA with Dunnett's post-hoc analysis. Data are pooled from two (A-B) or three (C-E) independent experiments with three replicates per condition.

Novel Corynebacterium secreted factor inhibits *S. aureus* hemolysis

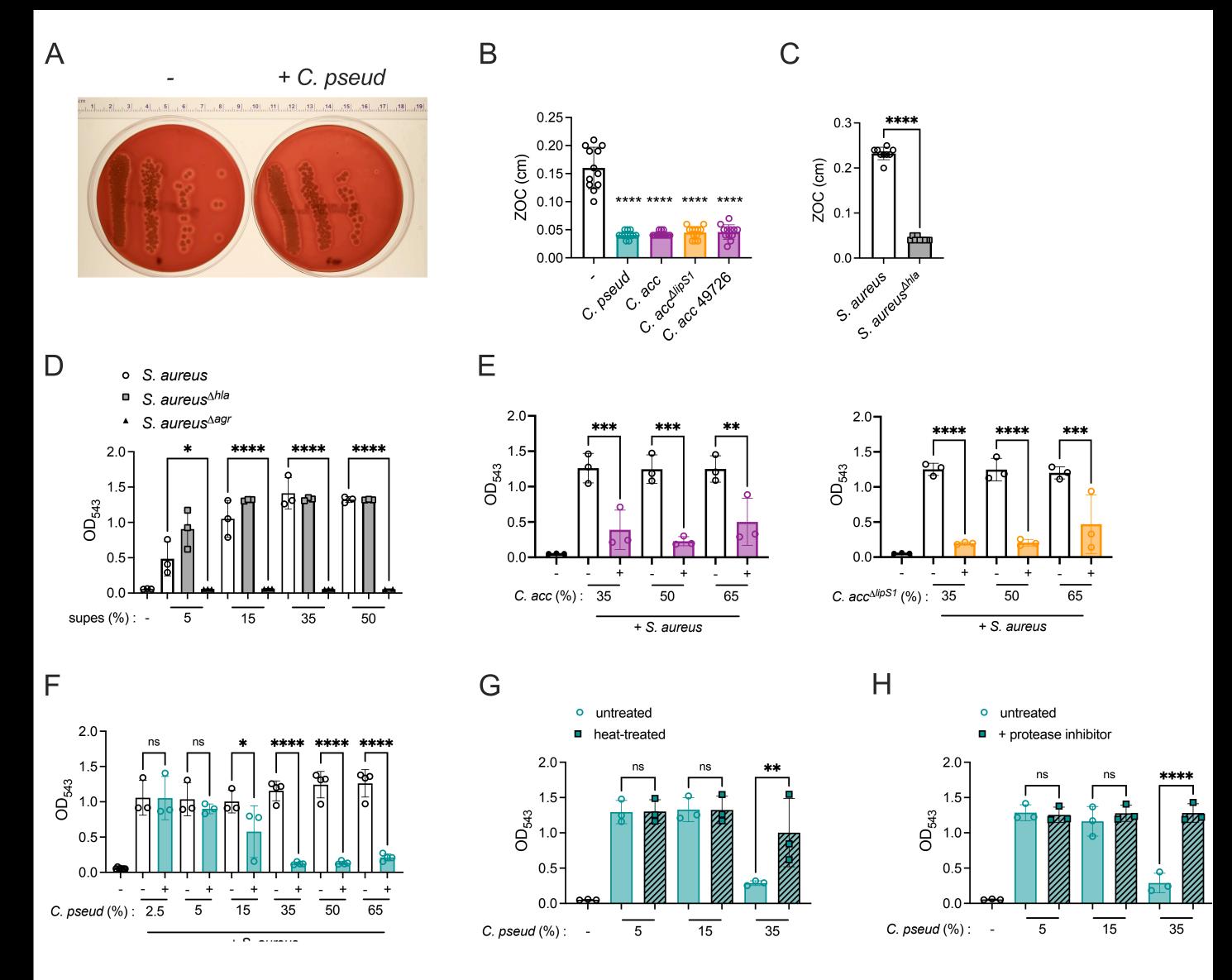


Figure 6. Corynebacterium secreted factor directly inhibits *S. aureus* hemolysin activity. (A) *S. aureus* colonies on sheep blood agar plates with or without pre-spreading plates with filtered supernatants (CFCM) from *Corynebacterium*, with hemolysis visualized as cleared zones (ZOC) surrounding colonies. (B) Zone of clearance (ZOC) quantified for *S. aureus* as tested as in (A). (C) ZOC quantified for WT *S. aureus* compared with *hla* deficient *S. aureus* as for (B). (D) Hemolysis detected as OD₅₄₃ of human red blood cells combined with the indicated percentage of filtered *S. aureus* supernatants. (E,F) Hemolysis as for (D) with addition of filtered supernatants (CFCM) from *Corynebacterium* at indicated percentages. (G,H) Hemolysis of human red blood cells as in (F) for *S. aureus* supernatants combined with untreated, heat-treated, or protease inhibited CFCM from *C. pseudodiphtheriticum*. *p<.05, **p<.01, ***p<.001, ****p<.001, ****p<.0001, one-way ANOVA with Dunnett's post-hoc analysis (A), Tukey's post-hoc analysis (D), or Sidak's post-hoc analysis (E-H), or unpaired t test (B). Data are representative of three experiments (A), pooled from three independent experiments with three replicates per group (B) or pooled from 3-4 independent experiments with one replicate per group (E-H).

Conclusions and Future Directions

Conclusions

- Pre-exposure to *C. pseudodiphtheriticum* displays dual protective effects against respiratory tract infection with *S. pnuemoniae* and *S. aureus*.
- Corynebacterium exhibit free fatty acid dependent inhibition of *S. pneumoniae*, highlighting the importance of including exogenous lipids in future testing models.
- *C. pseudodiphtheriticum* and *C. accolens* pre-colonization reduces *S. aureus* and *S. pneumoniae* adherence to epithelial cells in a lipase-independent manner which requires live bacteria, suggesting that a resident microbial population with *Corynebacterium* may offer protective effects against bacterial pathogen infection.
- Corynebacterium may reduce S. aureus virulence by blocking hemolytic activity through secretion of an unidentified novel factor.

Future Directions

 Future experimentation using protein purification techniques seeks to further categorize the secreted factor which Corynebacterium use to inhibit S. aureus hemolytic activity.

Acknowledgments

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