

DREADD-ing Stress: Using Chemogenetics to Bypass Variability and Go Right to the Source with CRF Activation



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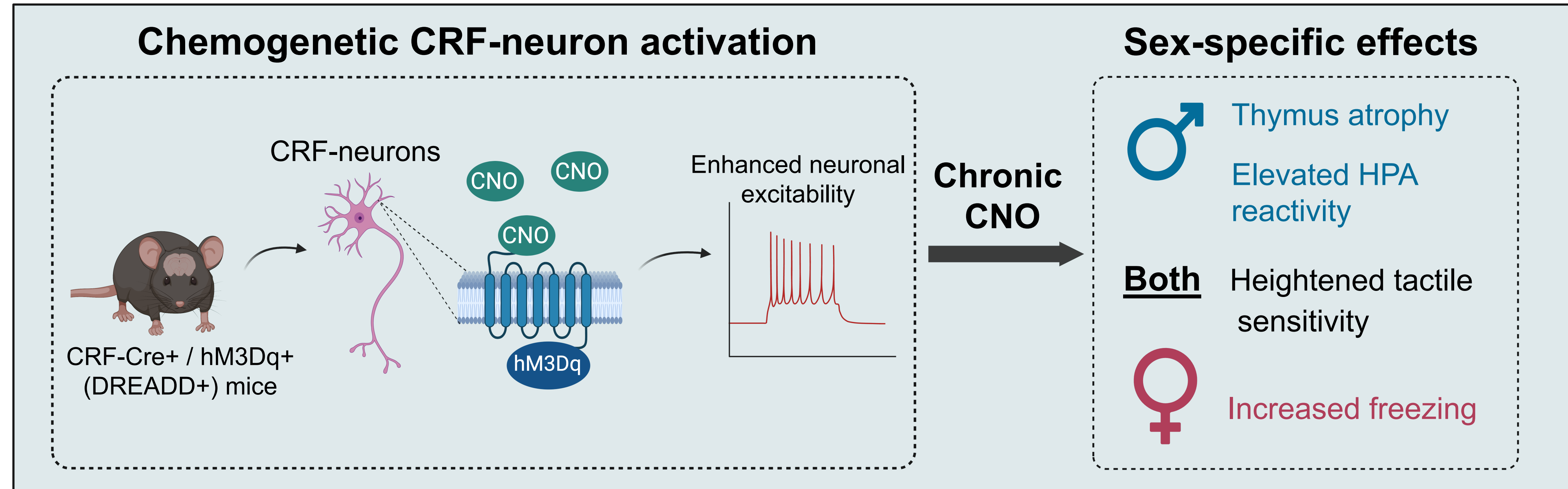
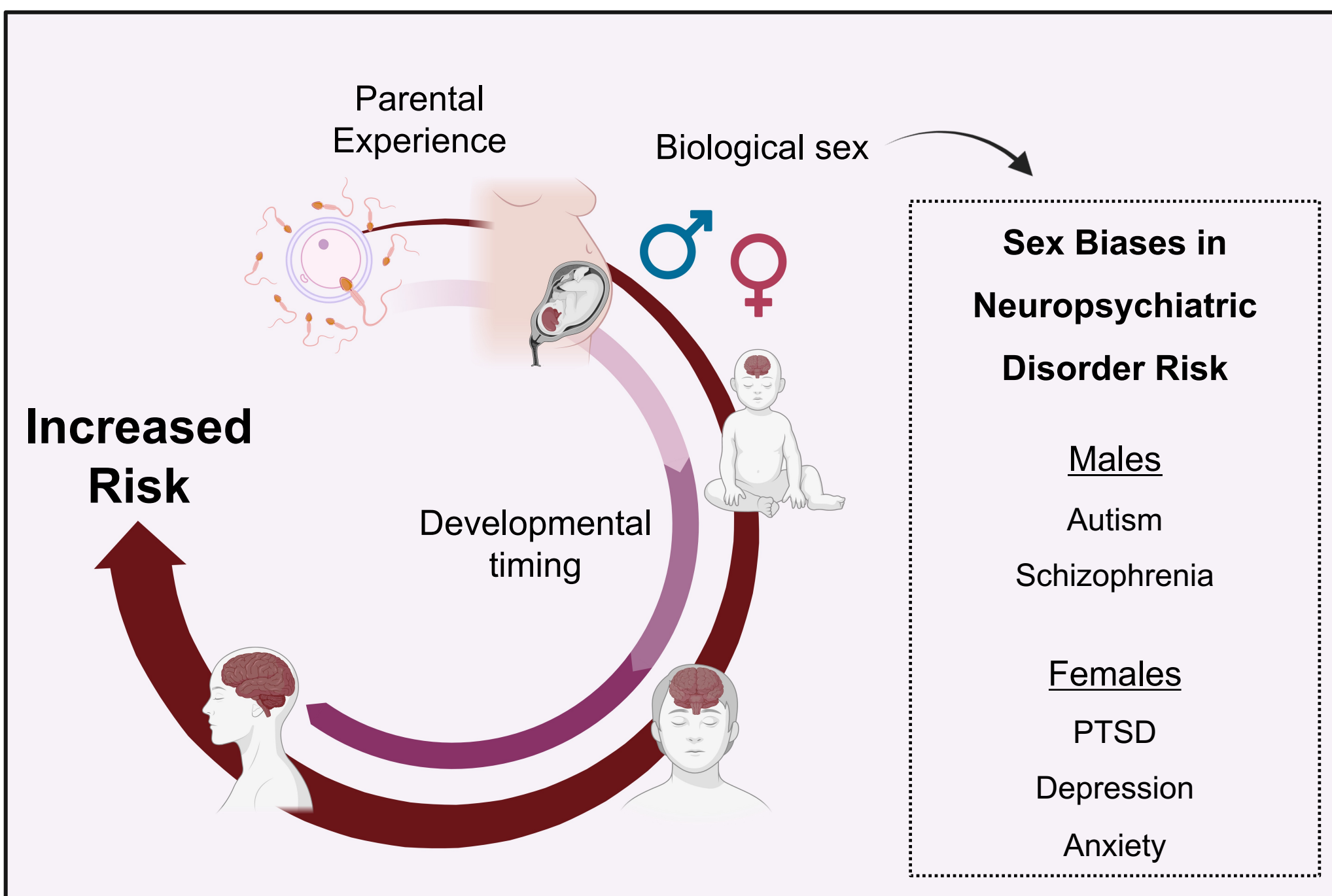
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Chronic stress across the lifespan is one of the strongest predictors of developing neuropsychiatric disorders. Risk is compounded by additional biological and environmental factors, including sex.

Existing stress models vary widely across research groups in stressor choice and exposure timing. Current paradigms use sensory stressors and require perception and experience of the stress. We hypothesized that repeated chemogenetic activation of CRF-neurons would mimic the physiological effects of chronic stress while bypassing the variability of experience and perception. We found that chronic CRF activation has sex-specific outcomes with males having increased sensitivity to the physiological effects and females showing greater limbic disruption.

Conclusions and Future Directions

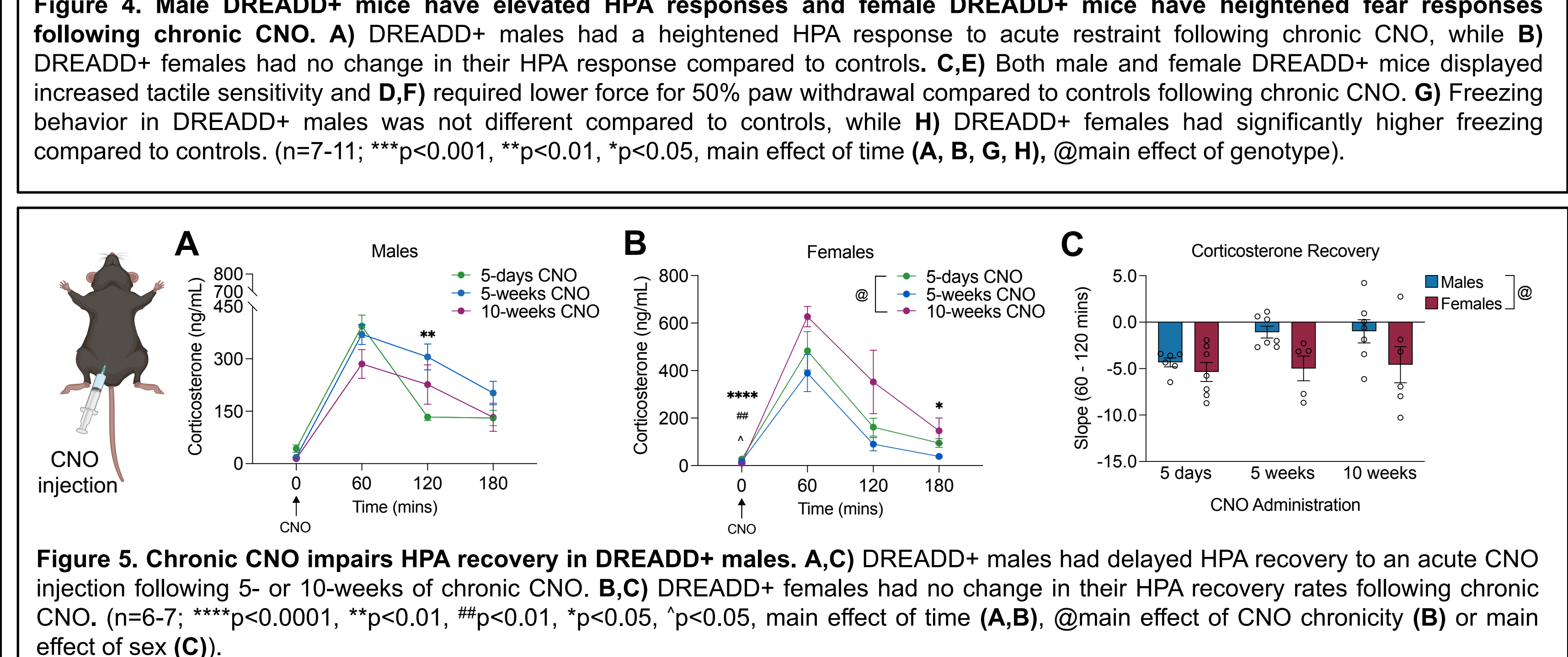
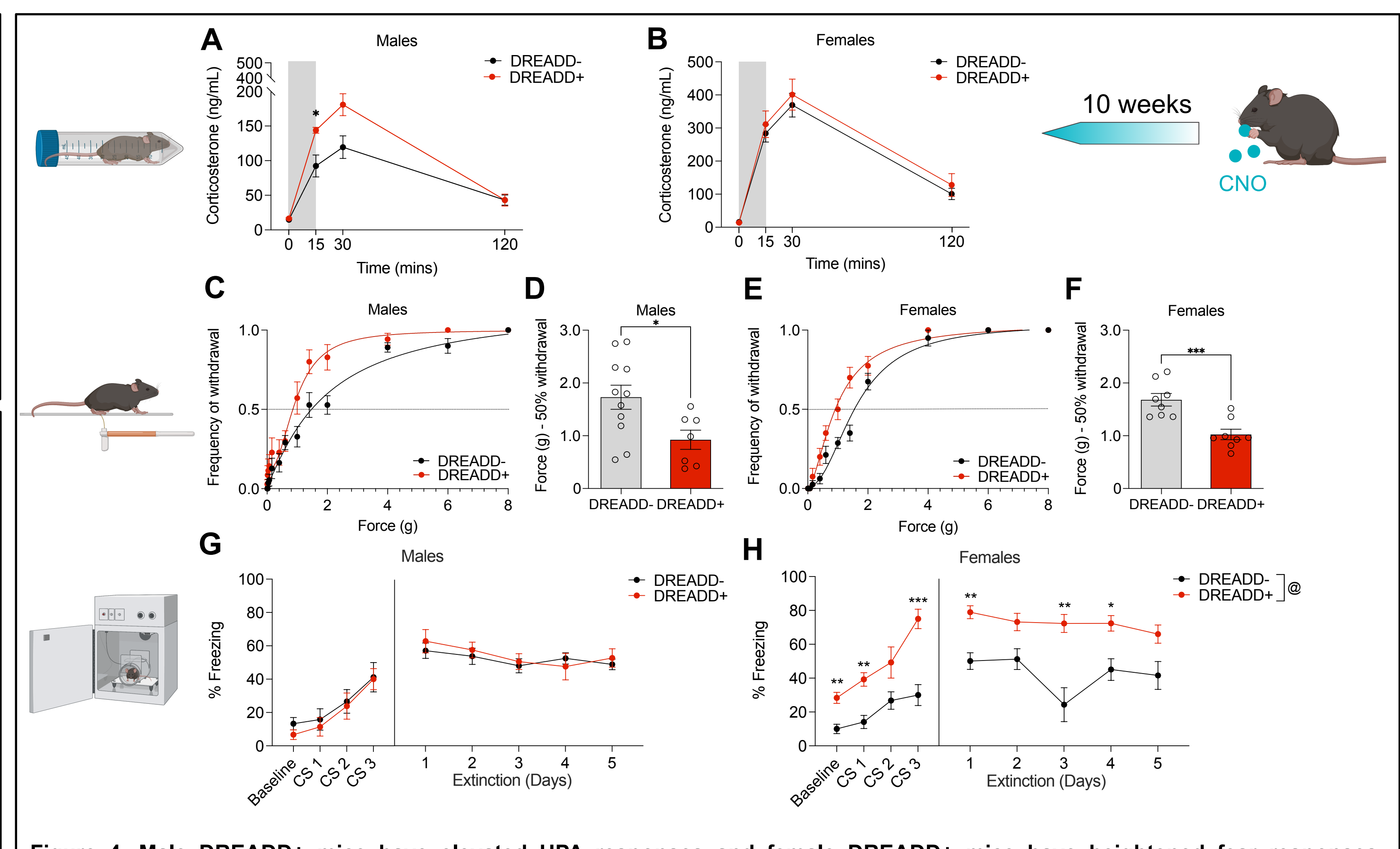
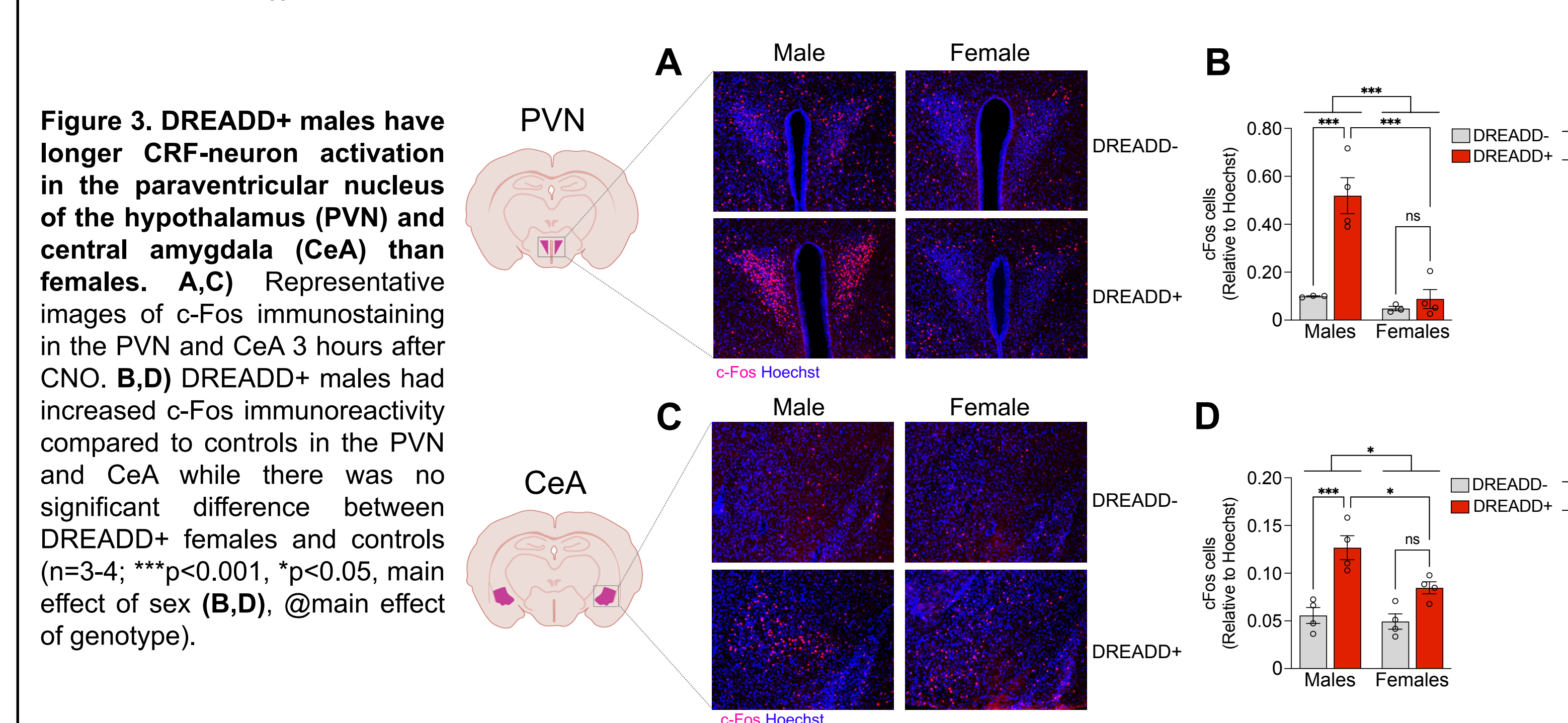
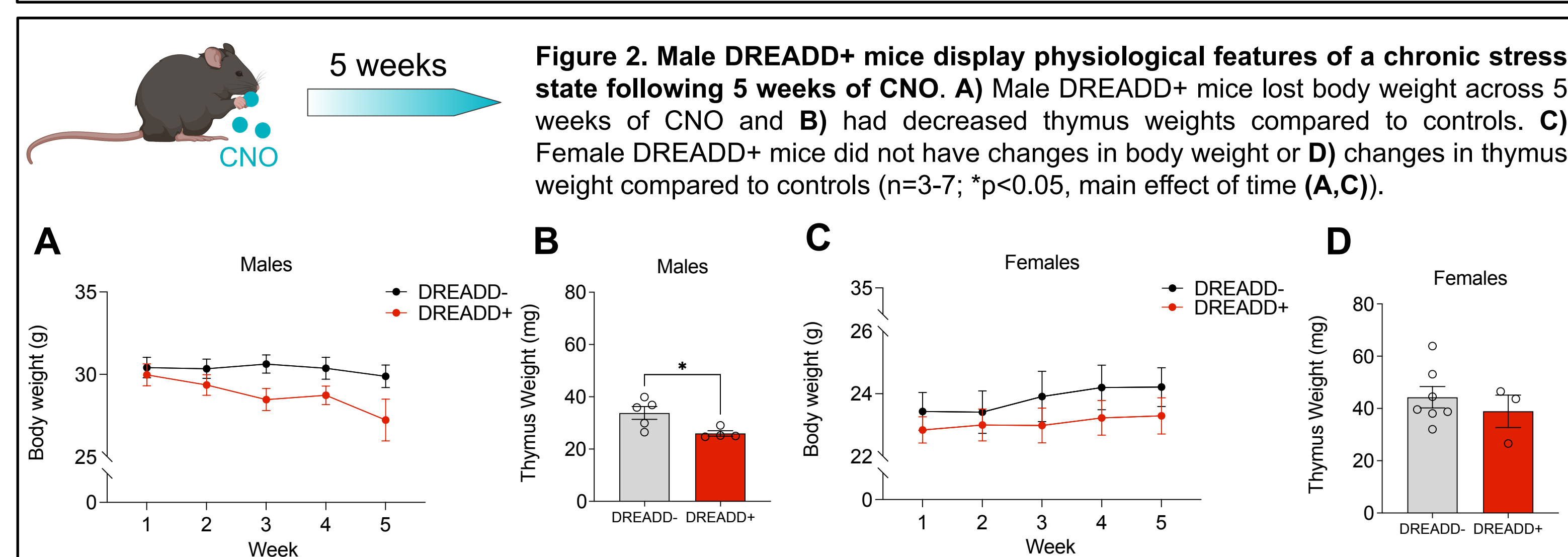
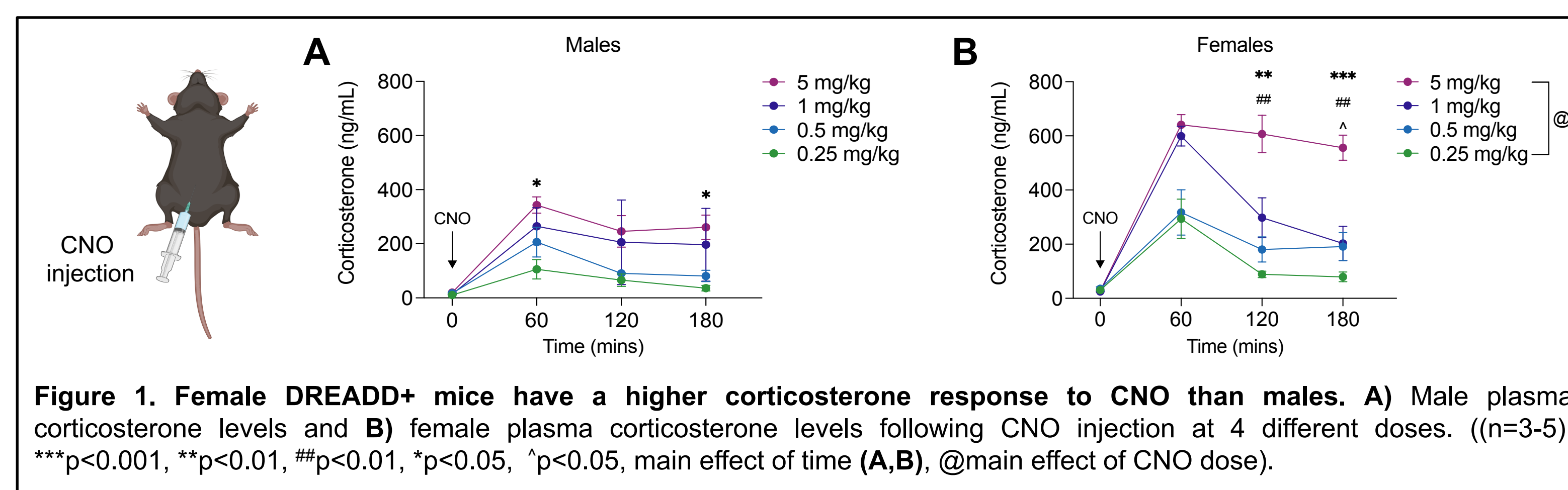
- Outcomes resulting from chronic stress are not solely due to increased glucocorticoid exposure.
- CRF-neurons in the PVN and CeA may be differentially sensitive to both chronic activation and sex, which could underlie the sex-biased presentation of many stress-related neuropsychiatric disorders.
- Future studies will examine the mechanisms underlying the sex-specificity and their developmental origins.



Animals: Homozygous CRF-Cre (Jax, stock #012704) mice were bred with heterozygous hM3Dq-DREADD (Jax, stock #026220) mice to generate CRF-Cre+/hM3Dq- (DREADD-) or CRF-Cre+/hM3Dq+ (DREADD+) offspring. Adult (10-18 week) animals were used for all studies. Mice were singly housed for the duration of experiments. **Clozapine-N-oxide (CNO):** CNO (HelloBio) was prepared at 1 mM in 0.9% normal saline and administered via intraperitoneal injection or cookie dough treat. **Hypothalamic-pituitary-adrenal (HPA) axis assessment:** Tail blood was collected from a small nick at the end of the tail at the specified timepoints. Plasma corticosterone levels were measured by radioimmunoassay (MP Biomedicals). **Immunohistochemistry:** Animals were transcardially perfused with ice-cold phosphate buffered saline followed by ice-cold 4% paraformaldehyde. Brains were stored in 4% paraformaldehyde for 24 hours then transferred to 30% sucrose for 24 hours at 4°C. Brains were frozen on dry-ice and stored at -80°C. 40 μ m sections were incubated overnight with anti-c-Fos (1:2500, Synaptic Systems) followed by goat-anti-guinea pig-Alexa Fluor 588 (1:200, Invitrogen) for 1 hour. **Imaging:** Imaging was performed using an ECHO-Revolve fluorescent microscope at 10X magnification. Images were analyzed using FIJI. **Von Frey Filament Test:** Following a 10-minute habituation, mice were placed on a suspended wire mesh and monofilaments of increasing diameter with forces ranging from 0.008 to 11.0 g (NC Medical) were pressed against the hind paw skin. Responses were recorded until the foot was withdrawn for 5 consecutive trials. **Fear Conditioning:** On day 1, mice were habituated to a chamber with an non-restrictive acrylic cylinder in a sound-attenuated box (context A, SR-LAB-Startle Response System, San Diego Instruments) for 10 minutes followed by the chamber in context B (original chamber modified with blue light covers, a black and white checkered pattern covering all walls, and 5% vinegar to clean surfaces) for 10 minutes. On day 2, the animals were placed in context A for 5 minutes and a 30 second baseline was collected with a 65 dB tone. The conditioned stimulus (CS; tone: 80 dB) was presented for 30 seconds followed by the CS paired with an unconditioned stimulus (US) of a 1s 0.6 mA foot shock 3 times. Fear memory extinction was measured on days 3-7. Mice were placed in context B and following a 5-minute acclimation period, a 30s baseline with the background tone was collected. The CS tone was presented for 30 seconds and the baseline-CS tone presentation was repeated for 15 trials with 30s intertrial intervals. Movement was measured using a piezoelectric accelerometer and recorded with SR-Lab software.

Figures were made with BioRender.com and GraphPad Prism. Statistical analysis was performed with GraphPad Prism.

Results



National Institute
of Mental Health



Eunice Kennedy Shriver National Institute
of Child Health and Human Development

These studies were supported by NIH HD097093 and MH108286 to TLB.