

# BURN INJURY, ALCOHOL, AND GUT ORGANOID GENE EXPRESSION

## ABSTRACT

Burn injuries remain a major source of morbidity and mortality worldwide and present clinicians with a complex set of challenges related to wound management, systemic inflammation, and multi-organ dysfunction. A striking epidemiologic feature of burn trauma is the high frequency with which alcohol intoxication co-occurs at the time of injury. Nearly half of burn patients in the United States present with measurable blood alcohol concentrations, and numerous studies have demonstrated significantly worse outcomes in this population [1,2]. Despite consistent clinical associations, the mechanisms linking alcohol intoxication to worsened burn outcomes remain incompletely understood.

Growing attention has been directed toward the gastrointestinal tract as a key driver of post-burn morbidity. The intestinal epithelium forms a dynamic barrier that protects against microbial translocation and contributes to systemic immune homeostasis. Burn injury disrupts this barrier through epithelial apoptosis, breakdown of tight junctions, impaired epithelial regeneration, and altered innate immune signaling [3,4]. Alcohol independently compromises the intestinal barrier by altering the mucus layer, increasing permeability, impairing antimicrobial peptide production, and promoting intestinal dysbiosis [5,6]. When burn injury and alcohol exposure occur together, the resulting impairment in intestinal defense is often greater than either insult alone [7,8].

Interleukin-22 (IL-22) is a cytokine crucial for coordinating epithelial regeneration and antimicrobial defense. IL-22 induces antimicrobial peptides such as Regenerating islet-derived protein 3 gamma (Reg3 $\gamma$ ) and membrane-bound mucins such as Mucin-1 (Muc1), both essential for maintaining intestinal barrier integrity [9–13]. Reg3 $\gamma$  is a C-type lectin that restricts microbial proximity to the epithelial surface, while Muc1 contributes to epithelial protection through anti-adhesive and immune-modulatory functions.

This study used a murine burn model paired with ex vivo colonic organoid cultures to investigate how ethanol exposure affects IL-22-mediated expression of Reg3 $\gamma$  and Muc1 following burn injury. Organoids were cultured from colonic crypts isolated from sham-injured and burn-injured mice, exposed to ethanol or vehicle, stimulated with IL-22, and analyzed using quantitative RT-PCR.

IL-22 robustly induced Reg3 $\gamma$  and Muc1 expression under all conditions. Ethanol significantly suppressed IL-22-induced Reg3 $\gamma$  expression in organoids derived from burn-injured mice, producing a 68 percent reduction compared with IL-22 stimulation alone. This effect was not observed in sham-derived organoids. Muc1 expression remained IL-22-dependent and unaffected by burn injury or ethanol exposure. These findings highlight selective vulnerability within IL-22-responsive antimicrobial pathways following combined burn injury and ethanol exposure.

Understanding how trauma and alcohol reshape epithelial antimicrobial responses is essential for designing interventions to reduce infection risk and systemic inflammation in intoxicated burn patients. This study demonstrates that combined burn injury and ethanol exposure impair IL-22-mediated Reg3 $\gamma$  expression but preserve Muc1 induction, revealing divergent responses among epithelial defense pathways and shedding light on mechanisms underlying increased susceptibility to complications in intoxicated burn patients.

## REFERENCES

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