

# INHIBITORY EFFECT OF CRYSTALLINS ON LENS EPITHELIAL TO MESENCHYMAL TRANSITION VIA BLOCKING OF $\alpha$ B-CRYSTALLIN ACTIVITY

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Cataract surgery is one of the most common surgeries in the world, with a little over two million surgeries performed annually in the United States alone. The most common complication is the development of a posterior capsular opacification (PCO), which occurs in over 20% of cases within 2-5 years after surgery. It has been shown that PCOs develop due to residual lens epithelial cells undergoing epithelial to mesenchymal transition (EMT). This can be instigated by TGF- $\beta$ 2, a predominant cytokine in the eye. EMT triggers the formation of fibrotic tissues and wrinkles in the posterior capsule. Our goal is to identify a potential agent that can prevent EMT via interactions with  $\alpha$ B-crystallin, a promoter of TGF- $\beta$ 2 signaling during the development of PCO. In this study, we aim to determine a mechanism by which two drugs, which we dubbed crystallin-1 and crystallin-2, affect the EMT process and how they specifically inhibit  $\alpha$ B-crystallin's behavior.

To determine whether the drugs inhibit  $\alpha$ B-crystallin chaperone activity, we assessed the chaperone activity using alcohol dehydrogenase (ADH) as a client protein in a thermal aggregation assay. The drugs at 20  $\mu$ M were incubated for four hours with  $\alpha$ B-crystallin, before we measured the ADH scattering at 400nm. For cell culture, we used the FHL124 cell line (fetal human epithelial cells) to determine the potency of the drugs in inhibiting the TGF- $\beta$ 2 induced EMT. Cells were pre-treated with 2  $\mu$ M of either drug for four hours, and then incubated with 10 ng/mL of human recombinant TGF- $\beta$ 2 for an additional 1-48 hours. Western blots were performed to measure the expression of TGF- $\beta$ 2 induced markers,  $\alpha$ SMA and  $\alpha$ B-crystallin. To assess effects on the TGF- $\beta$ 2 signaling pathway, we also measured the phosphorylation of SMAD2, AKT, and ERK.

The ADH aggregation assay revealed that both drugs significantly inhibited  $\alpha$ B-crystallin chaperone activity. Western blot data demonstrated that both drugs decreased the expression of  $\alpha$ SMA, but not the expression of  $\alpha$ B-crystallin; both of which are induced by TGF- $\beta$ 2 in FHL124 cells. Crystallin-2 was more efficacious in inhibiting EMT response than crystallin-1. In addition, neither drug influenced the phosphorylation of SMAD2, ERK, or AKT.

Our results suggest that both crystallins inhibit the TGF- $\beta$ 2 induced EMT of lens epithelial cells, but they do not alter the expression of  $\alpha$ B-crystallin. On the other hand, the drugs inhibit the chaperone activity of  $\alpha$ B-crystallin, therefore inhibiting EMT of lens epithelial cells. The lack of change in the phosphorylation of SMAD2, ERK, and AKT suggest that the crystallins do not influence either the canonical or the non-canonical signaling pathways. Taken together, crystallin-1 and crystallin-2 have a strong potential for use in the prevention of PCO.

Cataract surgery is one of the most common surgeries in the world, with a little over two million surgeries performed annually in the United States alone. The most common complication is the development of a posterior capsular opacification (PCO), which occurs in over 20% of cases within 2-5 years after surgery. It has been shown that PCOs develop due to residual lens epithelial cells undergoing epithelial to mesenchymal transition (EMT). This can be instigated by TGF-2, a predominant cytokine in the eye. EMT triggers the formation of fibrotic tissues and wrinkles in the posterior capsule. Our goal is to identify a potential agent that can prevent EMT via interactions with alphaB-crystallin, a promoter of TGF-beta2 signaling during the development of PCO. In this study, we aim to determine a mechanism by which two drugs, which we dubbed crystalatin-1 and crystalatin-2, affect the EMT process and how they specifically inhibit alphaB-crystallin's behavior.

To determine whether the drugs inhibit alphaB-crystallin chaperone activity, we assessed the chaperone activity using alcohol dehydrogenase (ADH) as a client protein in a thermal aggregation assay. The drugs at 20 microM were incubated for four hours with  $\alpha$ B-crystallin, before we measured the ADH scattering at 400nm. For cell culture, we used the FHL124 cell line (fetal human epithelial cells) to determine the potency of the drugs in inhibiting the TGF-beta2 induced EMT. Cells were pre-treated with 2 microM of either drug for four hours, and then incubated with 10 ng/mL of human recombinant TGF-beta2 for an additional 1-48 hours. Western blots were performed to measure the expression of TGF-beta2 induced markers, alpha-SMA and alpha-B-crystallin. To assess effects on the TGF-beta2 signaling pathway, we also measured the phosphorylation of SMAD2, AKT, and ERK.

The ADH aggregation assay revealed that both drugs significantly inhibited alpha-B-crystallin chaperone activity. Western blot data demonstrated that both drugs decreased the expression of alphaSMA, but not the expression of alphaB-crystallin; both of which are induced by TGF-beta 2 in FHL124 cells. Crystalatin-2 was more efficacious in inhibiting EMT response than crystalatin-1. In addition, neither drug influenced the phosphorylation of SMAD2, ERK, or AKT.

Our results suggest that both crystalatins inhibit the TGF-beta2 induced EMT of lens epithelial cells, but they do not alter the expression of alpha-B-crystallin. On the other hand, the drugs inhibit the chaperone activity of alpha-B-crystallin, therefore inhibiting EMT of lens epithelial cells. The lack of change in the phosphorylation of SMAD2, ERK, and AKT suggest that the crystalatins do not influence either the canonical or the non-canonical signaling pathways. Taken together, crystalatin-1 and crystalatin-2 have a strong potential for use in the prevention of PCO.