



39th MD-PhD

National Student Conference
July 12th-14th, 2024

Conference Program

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Welcome

Welcome to the 39th Annual National MD/PhD Student Conference! The National MD-PhD Student Conference brings together students, faculty, and alumni from over 60 institutions across the country to interact with and learn from each other in beautiful Breckenridge, Colorado. At the conference, attendees engage in discussions with current and aspiring physician scientists about current scientific developments and explore career issues specific to the MD-PhD program. ***This event is almost entirely student run and organized***; the planning and organizing committee each year is made up of a small group of current MD-PhD students at the University of Colorado.

We would like to thank our Keynote speakers – Frances Collins, MD/PhD, W. Kimryn Rathmell, MD/PhD, Michael Brenner, MD, and James DiCarlo, MD/PhD – for making the journey to share their work with us. We are also grateful to the discussion leaders and PSTP directors for taking the time to lead discussions and share their insights with us.

We hope that you gain insights into your own research and career as you participate in the scientific and career-oriented portions of the conference, as well as make certain to enjoy the surroundings of the Rocky Mountains. Please note that the most up to date information on the conference can be found on the Webex app, with additional information found on the website (<https://medschool.cuanschutz.edu/mstp/md-phd-conference>). If you need anything during your time here, please do not hesitate to ask one of the conference organizers.

Sincerely,

The 2024 Organizing Committee

Meet Your Student Planning Committee

The planning committee each year is largely run by a small group of volunteer MD-PhD students at the University of Colorado! All the members participated in activities related to conference planning, with each member spearheading specific tasks.

**Rachel Cohen, GS2: Co-Chair, Keynote
Speaker Coordinator, Communication
Liaison, Breckenridge Resort Liaison**

Studies the role of HIF and intraepithelial lymphocytes in the colonic epithelium.



**Jackson Stocking, GS2: Co-Chair,
Breckenridge Resort Liaison, Keynote
Speaker Coordinator**

Studies the cerebellar nuclei as a potential target of neuromodulation to treat movement disorders.



**Erin Fish, GS2: PSTP Showcase Liaison,
Poster Session Coordinator**

Studies mechanisms dictating dengue virus clearance from blood circulation.



**Ira Fleming, GS2: Website and Registration
Manager, Webex Platform Manager**

Studies vaccine persistence in the lymph node using novel molecular tools.



**Brandon Hilliard, GS2: PSTP Showcase
Liaison, DEI and Ethics Organizer**

Studies B cell peripheral tolerance and anergy in the context of autoimmunity.



**Jessica Beynor, GS2, Fundraising Co-Chair,
Keynote Speaker Coordinator, Webex
Facilitator**

Studies the role of estrogen in the HNSCC tumor microenvironment in response to radiation and immunotherapy treatment.



**Daniel Moskop, GS2, Communication
Liaison; Poster Session Coordinator**

Studies the impact of inflammation on therapy response in MDS.



**Stefano Ginocchio, GS2, DEI Event
Coordinator**

Studies impact of tissue stiffness in impaired angiogenesis in severe, early-onset fetal growth restriction.



Ashlyn Stahly, GS2, Fundraising Co-Chair

**Studies the dynamics and determinants of
tRNA repair.**



Breckenridge Resort Map



Conference Schedule

All times listed in **Mountain Standard Time**

Friday July 12th

| Time | Event | Location |
|--------------------|-----------------------------------------------------------------------------------|------------------------------------------------------|
| 9:30 AM – 12:30 PM | Registration | Atrium |
| 12:30 PM – 2:00 PM | Lunch Keynote Speaker: Dr. Rathmell | Elevation Ballroom |
| 2:00 PM – 3:00 PM | Oral Presentations: Immunology & Microbiology General Medicine & Physiology | Elevation Ballroom Forest |
| 3:00 PM – 3:30 PM | Coffee Break | Atrium |
| 3:30 PM – 4:30 PM | DEI / Ethics (Session A) | DEI: Elevation Ballroom Ethics: Blue Spruce/Aspen |
| 4:45 PM – 5:45 PM | Career Panel | Elevation Ballroom |
| 5:45 PM – 7:15 PM | Dinner Keynote Speaker: Dr. DiCarlo | Elevation Ballroom |
| 7:15 PM – 7:30 PM | Poster set up | Forest/Atrium |
| 7:30 PM – 9:00 PM | Poster Session I & Happy Hour | Forest/Atrium |

Saturday July 13th

| Time | Event | Location |
|--------------------|-----------------------------------------------|------------------------------------------------------|
| 7:30 AM – 9:00 AM | Breakfast | Elevation Ballroom |
| 8:30 AM – 12:00 PM | Funday Activities | Various |
| 11:30 AM – 2:00 PM | Lunch Career Panel & Networking Session | Elevation Ballroom |
| 2:00 PM – 3:00 PM | Keynote: Dr. Brenner | Elevation Ballroom |
| 3:00 PM – 3:30 PM | ASPA Presentation | Elevation Ballroom |
| 3:30 PM – 4:00 PM | Coffee Break | Atrium |
| 4:00 PM – 5:00 PM | DEI / Ethics (Session B) | DEI: Blue Spruce/Aspen Ethics: Elevation Ballroom |
| 5:15 PM – 6:30 PM | Poster Session II | Forest |
| 6:30 PM – 8:30 PM | Dinner Keynote Speaker: Dr. Collins | Elevation Ballroom |

Sunday July 14th

| Time | Event | Location |
|---------------------|---------------------------------------------------------------------------|-----------------------------------------|
| 8:30 AM – 9:00 AM | Breakfast | Elevation Ballroom |
| 9:30 AM – 10:30 AM | Oral Presentations: Bioengineering & Medical Imaging Cancer Biology | Blue Spruce/Aspen Elevation Ballroom |
| 10:45 AM – 11:45 AM | Oral Presentations: Neuroscience Molecular Biology & Pharmacology | Blue Spruce/Aspen Elevation Ballroom |
| 12:00 PM – 1:00 PM | Lunch Closing Remarks | Elevation Ballroom |

Keynote Speakers

Dr. Francis S. Collins, MD-PhD

NIH Distinguished Investigator in the intramural program of the National Human Genome Research Institute



Francis S. Collins, M.D., Ph.D., currently serves as an NIH Distinguished Investigator in the intramural program of the National Human Genome Research Institute, pursuing genomics research on type 2 diabetes and a rare disorder of premature aging called progeria. Dr. Collins is a physician-geneticist noted for his landmark discoveries of disease genes and his previous leadership of the international Human Genome Project, which culminated in April 2003 with the completion of a finished sequence of the human DNA instruction book. He served as director of the National Human Genome Research Institute at NIH from 1993-2008.

Dr. Collins then served as the 16th Director of the National Institutes of Health (NIH), appointed by President Barack Obama and confirmed by the Senate in 2009. In 2017, President Donald Trump asked Dr. Collins to continue to serve as the NIH Director. President Joe Biden did the same in 2021. For those 12 years, serving an unprecedented three administrations, Dr. Collins oversaw the work of the largest supporter of biomedical research in the world, spanning the spectrum from basic to clinical research. Dr. Collins stepped down as NIH Director on December 19, 2021.

From February 2022 to October 2022, Dr. Collins served as Acting Science Advisor to President Biden. From November 2022 to May 2023 he continued his White House service as a Special

Advisor to the President for Special Projects. He continues to serve as the White House lead for a bold program to eliminate hepatitis C in the United States.

Dr. Collins is an elected member of both the National Academy of Medicine and the National Academy of Sciences, was awarded the Presidential Medal of Freedom in November 2007, and received the National Medal of Science in 2009. In 2020, he was elected as a Foreign Member of the Royal Society (UK) and was also named the 50th winner of the Templeton Prize, which celebrates scientific and spiritual curiosity.

Dr. W. Kimryn Rathmell, MD-PhD

Director of the National Cancer Institute

W. Kimryn Rathmell, M.D., Ph.D., M.M.H.C., was sworn in as the 17th NCI director on December 18, 2023. She previously led the Vanderbilt University Medical Center as physician-in-chief and chair of the Department of Medicine.



Dr. Rathmell is a recipient of the 2019 Louisa Nelson Award for Women of Achievement, Vision, and Inspiration the 2019 Eugene P. Schonfeld Award from the Kidney Cancer Association, and the Paragon Award for Research Excellence from the Doris Duke Foundation. She was a leader of The Cancer Genome Atlas's (TCGA) kidney cancer projects and served as a TCGA analysis working group member across the spectrum of cancers, winning the 2020 American Association for Cancer Research Team Science Award. She has served on the NCI Board of Scientific Advisors, and the Forbeck Foundation Scientific Advisory Board.

Dr. Rathmell has held leadership positions with the American Society of Clinical Oncology and the American Society for Clinical Investigation, serving as secretary-treasurer and president. As a result of her efforts, Dr. Rathmell has been elected to the Association of American Physicians, the American Academy of Arts and Sciences, and the National Academy of Medicine.

Dr. Rathmell's specialty is the research and treatment of complex and hereditary kidney cancers. She also focuses on underlying drivers of kidney cancers using genetic, molecular, and cell biology to develop interventions to improve patients' lives. Dr. Rathmell's research has resulted in more than 250 articles in leading peer-reviewed journals, including The New England Journal of Medicine, Nature, Proceedings of the National Academy of Sciences, and the Journal of Clinical Investigation.

Dr. Rathmell earned undergraduate degrees in biology and chemistry from the University of Northern Iowa and her Ph.D. in biophysics and M.D. from Stanford University. She completed an internal medicine internship at the University of Chicago and an internal

medicine residency, medical oncology fellowship, and postdoctoral studies at the University of Pennsylvania. In 2022, she completed her Master of Management in Health Care from the Vanderbilt University Owen Graduate School of Management.

Dr. Michael B. Brenner, MD

E. F. Brigham Professor of Medicine at Harvard Medical School, Director of the Human Immunology Center, Director of the Single Cell Genomics Core, and Director of Cell and Molecular Immunology at Brigham and Women's Hospital

Michael B Brenner MD is the E. F. Brigham Professor of Medicine, at Harvard Medical School and Director of the Human Immunology Center, Director of the Single Cell Genomics Core, and Director of Cell and Molecular Immunology at Brigham and Women's Hospital. Selected honors include: Election to the National Academy of Sciences and in 2023 named as Highly Cited Researcher by Clarivate (top 1% of scientists by citations).



Brenner's laboratory designs and implements high dimensional immunophenotyping, single cell transcriptomic analyses and functional studies to deconstruct human autoimmune disorders. Discovery-based single cell and spatial analyses carried out in humans are used to define pathologic cell populations which are then interrogated in vitro or in animal models. In recent studies in the T cell field, his laboratory defined a new T helper cell population called T peripheral helper T (Tph) cells that are pathologically expanded in rheumatoid arthritis, lupus and many other autoimmune diseases and drive B cell responses and antibody production. He recognized the major CD8 T cell population expanded in inflamed human tissues was not the well-known granzyme B+ cytotoxic T cells, but rather CD8 T cells that express granzyme K. Exciting new studies now reveal that granzyme K drives a new pathway of complement activation. He identified a super-activated macrophage state that produces the highest levels of inflammatory factors in inflamed synovium as well as in tissues across several diseases. His laboratory has defined the central role of fibroblasts in inflammation and tissue damage, including novel subsets of highly inflammatory fibroblasts expanded in RA, and the role of Notch3 signalling in driving fibroblast differentiation and pathologic function. Current studies are defining the transcriptional regulation of pathologic fibroblasts. Together, these studies are "reconstructing" the immunopathology of RA and other autoimmune diseases.

Some of Professor Brenner's other research accomplishments in basic biology include the discovery of gd T cells and their functions. In contrast to peptide MHC presentation, he identified a new system of antigen presentation by which immune system T cell receptors recognize lipids as cognate antigens presented by CD1 antigen presenting molecules. He defined the trafficking, loading and function of CD1a, b, c and d, and the functions of CD1-restricted T cells and iNKT cells in host defense and immunoregulation. He identified the integrin molecule αEb7, its counter-receptor (E-cadherin), and its role in mucosal leukocyte

homing. He defined the role of calnexin in protein assembly and quality control, and the role of small GTPase Arl proteins in endocytic trafficking.

Dr. James J. DiCarlo, MD-PhD

Peter de Florez Professor of Neuroscience Director, MIT Quest for Intelligence Co-Director, and Center for Brains, Minds, and Machines Investigator at McGovern Institute for Brain Research at Massachusetts Institute of Technology



Jim DiCarlo is a Professor of Systems and Computational Neuroscience at the Massachusetts Institute of Technology. His research team's primary goal is to discover and artificially emulate the brain mechanisms that underlie human visual intelligence. Over the past 20 years, using the non-human primate animal model organism, DiCarlo and his collaborators have helped develop our contemporary, engineering-level understanding of the neural mechanisms that underlie visual information processing in the ventral visual stream — a complex series of interconnected brains areas — and how that processing supports core cognitive abilities such as object and face recognition. He and his collaborators aim to use this newly emerging scientific understanding to guide the development of more robust artificial vision systems ("AI"), to reveal

new ways to beneficially modulate brain activity via modulations of images striking our eyes, to expose new methods of accelerating visual learning, to provide a basis for new neural prosthetics (brain-machine interfaces) to restore lost senses, and to provide a scientific foundation to understand how sensory processing is altered in conditions such as agnosia, autism and dyslexia.

DiCarlo trained in biomedical engineering, medicine, systems neurophysiology and computing at Northwestern (BSE), Johns Hopkins (MD/PhD), and Baylor College of Medicine (Postdoc). He served as Head of MIT's Department of Brain and Cognitive Sciences from 2012 to 2021, and he is currently the Director of the MIT Quest for Intelligence (2021-present) where he and his leadership team are working to advance interdisciplinary research at the interface of natural and artificial intelligence. DiCarlo is an Alfred P. Sloan Research Fellow, a Pew Scholar in Biomedical Sciences, a McKnight Scholar in Neuroscience, and an elected member of the American Academy of Arts & Sciences.

Panelists:

Diversity, Equity, and Inclusion: Dr. Cherise Hamblin, MD

Founder and President of Patients R Waiting, Assistant Professor in OBGYN at UMass Chan Medical School, Medical Director of the UMass Memorial Health Doula Program, Director of the Underrepresented in Medicine Workforce Development and Capacity Building for the UMass Chan Collaborative in Health Equity

Dr. Cherise Hamblin was born and raised in the Bronx, NY where she attended Bronx High School of Science. After graduation, Dr. Hamblin attended Franklin & Marshall College

where she received a double major in biology and Spanish in 2003. During her time at Franklin & Marshall College, she served as the Black Student Union Vice President and the Admission Office intern for Minority Recruitment. She also played rugby and studied abroad in Guadalajara, Mexico. Dr. Hamblin subsequently attended medical school at Northwestern University and completed her training in Obstetrics & Gynecology in Phoenix, AZ. Dr. Hamblin is passionate about working with the next generation of physicians both in her clinical roles and nonprofit work. For over a decade she has worked with advisors, students, and community organizers on various mentoring and health career exposure programs.



She is the founder and president of Patients R Waiting, a non-profit organization dedicated to eliminating health disparities by increasing diversity in medicine. She is a board certified OBGYN with a career focus on health care workforce development and antiracism in medicine. She is an Assistant Professor in OBGYN at UMass Chan Medical School and the Medical Director of the UMass Memorial Health Doula Program. Additionally, she serves as the Director of Underrepresented in Medicine Workforce Development and Capacity Building for the UMass Chan Collaborative in Health Equity.

Ethics Discussion: Dr. Insoo Hyun, PhD

Director of the Center for Life Sciences and Public Learning at the Museum of Science in Boston

Insoo Hyun, PhD, is a prominent figure in bioethics, with significant contributions to research ethics and policy in advanced biotechnologies. He is the former director of research ethics at Harvard Medical School's Center for Bioethics and taught at HMS as a senior lecturer in Global Health and Social Medicine. Currently, Dr. Hyun is the director of the Center for Life Sciences and Public Learning at the Museum of Science, Boston. He also works as a bioethics consultant for the Broad Institute of MIT and Harvard.

Dr. Hyun's expertise is widely recognized through his appearances on National Public Radio and contributions to journals like *Nature* and *Science*. He authored *Bioethics and the Future of Stem Cell Research* and was honored as one of Cleveland's Most Interesting People of 2019.



Career Panelists

| | |
|-----------------------------------------------|---------------------------------------------------------------------------------------------|
| | |
| Dr. W. Kimryn Rathmell, MD, PhD, MMMHC | National Cancer Institute Director |
| Dr. Charles Emala, MD | Columbia University Department of Anesthesiology Vice Chair for Research |
| Dr. Melanie Cree, MD, PhD | Pediatric Endocrinology at University of Colorado Anschutz and Children's Hospital Colorado |
| Dr. Taylor Soderborg, MD, PhD | Medical Director at Tiny Health |

PSTP Showcase

| The Ohio State University | Internal Medicine PSTP | Dr. Robert Baiocchi, MD, PhD |
|-------------------------------------------------|---------------------------------------------|-----------------------------------|
| Harvard University | Internal Medicine PSTP | Dr. Jatin Vyas, MD, PhD |
| University of California San Francisco | UCSF Physician Scientist Scholar Program | Dr. Arun Wiita, MD, PhD |
| University of North Carolina Chapel Hill | Pediatric PSTP | Dr. Misty Good, MD, MS |
| Columbia University | Anesthesiology Physician Scientist Research | Dr. Charles Emala, MD |
| Duke University | Anesthesiology PSTP | Dr. Mara Serbanescu |
| University of North Carolina Chapel Hill | Internal Medicine PSTP | Dr. Joseph (Alex) Duncan, MD, PhD |
| Stanford University | Anesthesiology PSTP (FARM Program) | Dr. QiLiang Chen, MD, PhD |
| Stanford University | Internal Medicine PSTP (TIPS Program) | Dr. Paul Cheng, MD, PhD |
| University of Colorado | Internal Medicine PSTP | Dr. David Schwartz MD |
| University of Colorado | Pediatric PSTP | Dr. Christine Vohwinkel MD, PhD |
| University of Michigan | Pathology PSTP | Dr. Mark Rudolf, MD, PhD |

Student Oral Presentation Schedule and Abstracts

Concurrent Session 1A

I. Immunology and Microbiology (Elevation Ballroom)

Host response during urinary tract infection alters mammary extracellular matrix and accelerates Brca1 tumorigenesis, Steven Lewis, Stony Brook University

Environmental factors are known to potentiate the tumorigenic potential of genetically predisposed cells. While breast cancer is among the most common tumors in women and many genetic risk factors are known, there is a paucity of known environmental risk factors. Our recent work uncovered that an environmental factor, urinary tract infection, through systemic inflammatory mediators induced during the host response can remodel the mammary gland at the cellular and tissue level. We found that wildtype mice with an unresolved UTI demonstrate ductal hyperplasia, fibroblast activation and collagen deposition, which depended on systemic TIMP1 induced by UTI. Because ECM changes can induce epithelial proliferation through the YAP pathway, which we found using scRNAseq was elevated in mammary epithelial cells during UTI, we hypothesized that the UTI host-response might impact breast tumorigenesis in a genetically predisposed host. In our autochthonous mouse model of Brca1-deficient breast cancer, we discovered that the systemic inflammatory response triggered by UTI promotes tumorigenesis. Through histological analysis of pre-tumor and tumor tissue, scRNA-seq and transplant experiments, we identified that the systemic response to UTI induced expansion of a Krt6a-expressing basal-luminal (BL) epithelial population with enhanced plasticity and tumor initiating potential. We traced the activation of these Krt6a+ basal-luminal cells through mechano-sensing gene transcription driven by YAP/Tea1 back to ECM deposition of collagen and tenascin-C by fibroblasts activated after UTI. These ECM changes engaged Itgb6 and induced proliferative signaling in BL cells. This collagen deposition and activation of the tumor initiating capacity of BL cells by UTI could be reversed through resolution of the infection using antibiotic treatment or TIMP1 neutralization. This finding provides the first example of how a local infection, commonly experienced by women globally, can impact the expressivity of a common cancer associated mutation in breast cancer and presents an avenue for improved prevention and surveillance efforts in the BRCA1 population.

B cell peripheral tolerance is promoted by cathepsin B protease, Marissa Chou, University of California San Francisco

B cells that bind soluble autoantigens receive chronic signaling via the B cell receptor (signal-1) in the absence of strong costimulatory signals (signal-2), and this leads to their elimination in peripheral tissues. The factors determining the extent of soluble autoantigen-binding B cell elimination are not fully understood. Here we demonstrate that elimination of B cells chronically

exposed to signal-1 is promoted by cathepsin B (Ctsb). Using hen egg lysozyme- (HEL) specific immunoglobulin transgenic (MD4) B cells and mice harboring circulating HEL, we found improved survival and increased proliferation of HEL-binding B cells in Ctsb-deficient mice. Bone marrow chimera experiments established that both hematopoietic and non-hematopoietic sources of Ctsb were sufficient to promote peripheral B cell deletion. Depletion of CD4⁺ T cells overcame the survival and growth advantage provided by Ctsb deficiency, as did blocking CD40L or removing CD40 from the chronically antigen-engaged B cells. Thus, we suggest that Ctsb acts extracellularly to reduce soluble autoantigen-binding B cell survival and that its actions restrain CD40L-dependent pro-survival effects. These findings identify a role for cell-extrinsic protease activity in establishing a peripheral self-tolerance checkpoint.

Exploring the role of fibroblast Reg3 proteins in response to intestinal inflammation, Sritejasvinthi Karimikonda, Medical College of Wisconsin

Fibroblasts are multifaceted cells involved in extracellular matrix formation, immune regulation, and tissue homeostasis. Fibroblasts proliferate in response to disease and are involved in the pathology of many inflammatory diseases including inflammatory bowel disease (IBD) by contributing to complications like tissue fibrosis and stricture formation. *However, the processes which drive fibroblast response to intestinal inflammation and the role of fibroblasts in host defense are still poorly understood.* To investigate the intestinal fibroblast response, we employ a bacterial enteritis model system. Using spatial transcriptomics, we found increased expression of *regenerating islet-derived protein beta (Reg3 β)* and *gamma (Reg3 γ)* in the cecum in response to infection. Only epithelial and gamma-delta T cells have been shown to express *Reg3 β* and *Reg3 γ* in the gut thus far. Flow cytometry analysis showed that in addition to epithelial cells and immune cells, an Epcam- Cd45- subset of cells, which includes fibroblasts, also produced *Reg3 γ* . *Fibroblast expression of Reg3 proteins has not been previously demonstrated.* Integration of our spatial transcriptomics dataset with a small intestine scRNAseq dataset showed that within the non-epithelial and non-immune cells subset, both fibroblasts and endothelial cells can express *Reg3 β* and *Reg3 γ* transcripts. We explored this further using the spatial transcriptomics data, and found an increased number of fibroblast marker transcripts, *Pdgfra*, *Thy1*, *Pdpn*, and *Vim*, co-localized with *Reg3 β* and *Reg3 γ* transcripts in response to intestinal inflammation. Moreover, there were more transcript co-localizations associated with fibroblast markers compared to endothelial cell markers, suggesting that fibroblasts are a key cell type involved in *Reg3* expression. Overall, our results suggest a novel fibroblast innate immune response to intestinal inflammation via antimicrobial peptide production.

Concurrent Session 1B

II. General Medicine and Physiology (Blue Spruce/Aspen)

Investigating the role of heme in acute and chronic sickle cell disease pain, Samuel Zorn, Medical College of Wisconsin

Individuals with sickle cell disease (SCD) suffer from complex stimulus-evoked and spontaneous pain that is mediated in part by the hyperexcitability of their peripheral sensory neurons. However, it is unclear whether acute or chronic elevation of cell free heme, a key pathological feature of SCD, contributes to the aberrant activity of sensory neurons. Thus, determining whether heme drives pain behavior and enhanced excitability of dorsal root ganglia (DRG) primary sensory neurons involved in pain signal transduction is critical for the development of targeted pain therapies for SCD. In the current experiments, we determine whether cell free heme accumulates within the DRG and further use evoked and ongoing pain behavior assays and *in vitro* calcium imaging and patch clamp electrophysiology to evaluate the hypothesis that heme causes direct sensitization of nociceptive DRG neurons in wildtype (WT) mice. Our preliminary data reveal that similar to pain behaviors in the mouse model of SCD, hind paw injection of heme into WT mice induces mechanical and cold hypersensitivity, place aversion, and facial grimace. These data suggest that sensory terminals in the skin become hyperexcitable when exposed to elevated heme. We next administered heme to primary cultured DRG neurons *in vitro* and observed calcium flux across the neuronal cell membrane that was dependent on extracellular calcium. Together, these data suggest that heme opens calcium channels on sensory neurons to drive acute pain behaviors. As ongoing activity of nociceptive neurons may cause pain signal transduction in the absence of external stimuli, we will next use current clamp electrophysiology to determine whether application of heme to cultured DRG neurons is sufficient to induce action potential firing. Finally, our preliminary data suggests that injection of exogenous haptoglobin, a hemoglobin scavenger that prevents heme accumulation, alleviates steady-state pain behaviors in Townes SCD mice. Completion of these experiments will shed light on the contributions of cell free heme to SCD pain and illuminate downstream effectors that may be targeted to provide analgesic relief for this historically understudied and debilitating disease.

Longitudinal blood pressure trajectories associated with exposure to metals and metal mixtures, Margaret Weiss, University of Illinois at Chicago

Background: Elevated blood pressure is a leading risk factor for cardiovascular disease, the leading cause of mortality worldwide. Evidence suggests that metal exposure may increase blood pressure; however, most previous studies have been cross-sectional or underpowered. Furthermore, underserved racial/ethnic groups are both at higher risk of hypertension and metal exposure, including Mexican Americans. Consequently, we aimed to evaluate associations between longitudinal blood pressure traits in relation to urinary levels of metals and metal mixtures among 431 Mexican American adults. **Methods:** A longitudinal cohort of 431 Mexican

American adults, ages 35 to 69, were followed over 3 years with 6 repeated examinations. Urinary metals were quantified at baseline. Linear mixed effects models were fitted to estimate the per month increase in systolic blood pressure, diastolic blood pressure, pulse pressure, and mean arterial pressure associated with higher urinary metal levels. Metal mixture scoring, principal component analysis, weighted quantile sum regression, and quantile-based g computation were utilized to assess the associations of the metal mixture. **Results:** After adjustment, single metal models identified arsenic, cobalt, molybdenum, and zinc with faster increases in blood pressure. Arsenic was associated with a faster per month increase in systolic blood pressure, pulse pressure, and mean arterial pressure; molybdenum with systolic and pulse pressure; and zinc and cobalt with pulse pressure. The metal mixture was associated with a faster per month increase in pulse pressure. **Conclusion:** This study identifies several metals, both individually and as mixtures, associated with increased blood pressure traits over time. Since elevated blood pressure is a driver of cardiovascular disease, for which incidence continues to disproportionately affect Hispanic/Latino individuals, this study provides insight into the impacts of metals on cardiometabolic health.

Trajectories of mHealth-Tracked Mental Health Symptoms and Their Predictors in Chronic Pelvic Pain Patients, Emily Leventhal, Icahn School of Medicine at Mount Sinai

Background: Female chronic pelvic pain disorders (CPPDs; e.g., endometriosis, adenomyosis, fibroids) are prevalent and debilitating conditions associated with co-morbid clinical anxiety and depressive symptoms. Characterizing variable inter-individual symptom trajectories and lifestyle factors as possible predictors is a starting point to enable patient self-efficacy and symptom management. We herein investigate the association between mental health, pain symptoms, and physical activity (PA) in CPPDs and demonstrate a method for handling multi-modal, non-linear mobile health (mHealth) data for this clinical scenario. **Methods:** Study sample included 3,640 person-level days of data from 62 CPPD patients. Participants completed the McGill Pain Questionnaire (MPQ), PROMIS pain interference (PI), pain self-efficacy (PSEQ), PROMIS global mental health (GMH) for 14 weeks using a research mHealth App, and the pain catastrophizing scale (PCS) and psychiatric history at baseline. We used activity trackers to collect daily moderate-to-vigorous PA (MVPA) data. **Data analysis:** We used generalized additive mixed-effects models (GAMMs) to regress weekly GMH scores on 7-day MVPA and other weekly pain outcomes and adjusted for baseline measures, time in study, and the random intercept of the individual. We converted 7-day MVPA data into a single smooth using spline basis functions with a polynomial term ($n=3$), to model the potential non-linear relationship. **Results:** MVPA was a significant, curvilinear predictor of GMH scores ($B_1=1.60$; $B_2=-7.07$, $B_3=14.10$, $p<0.05$), independent of psychiatric diagnoses and weekly pain-related factors. PI and PCS were negatively associated with GMH scores ($B=-2.41$, $B=-3.35$, respectively; $p<0.05$). **Conclusion:** Our findings underscore the complex, non-linear relationship between PA and mental health in CPPD and demonstrate the potential of ambulatory mHealth-based data combined with GAMMs for delineating inter-individual and temporal variability.

Concurrent Session 2A

I. Bioengineering and Medical Imaging (Blue Spruce/Aspen)

Rethinking Spinal Fusion Surgery: A new paradigm in the biology of spinal fusion, Doby Okonkwo, Vanderbilt School of Medicine

Introduction: Fusion of the spine is crucial for treating many spinal disorders that cause pain and dysfunction. Forced ossification between vertebra is essential for successful fusion, and failure rates remain high at 35%, even with supplemental bone grafts. The current paradigm posits that spinal fusion is driven by the differentiation of mesenchymal stem cells (MSCs) from the intraosseous bone directly into osteoblasts without a chondrocyte intermediate. This approach neglects the outer bone membrane, periosteum, which is removed to expose trabecular bone. Supported by recent findings in fracture repair, we propose a new paradigm: MSCs originating from the periosteum drive ossification through a chondrocyte intermediate. **Methods:** Advanced lineage tracing animals were used to determine the origin of MSCs and type of bone formation required for spinal fusion in a clinically relevant pre-clinical murine model. We also explored if different surgical techniques in regard to periosteal health affect fusion. Bone formation was assessed through radiographs, microCT analysis, and histology. **Results and Discussion:** Our findings reveal that MSCs originate from the periosteum for bone formation during spinal fusion, undergoing chondrogenesis as an intermediate step. Cauterizing the periosteum significantly reduced MSC recruitment, highlighting the vital role of the periosteum. **Conclusion:** Our study shifts the paradigm of the origin of MSCs required for spinal fusion from the intraosseous trabeculae to the outer membrane of bone, the periosteum. This shift has significant implications for spinal fusion surgery, which previously viewed the periosteum as a layer needed to remove to expose the trabecular bone. Additionally, our findings challenge the focus on promoting intramembranous ossification, suggesting that grafts which promote chondrogenesis, as opposed to osteogenesis, improve clinical results.

Polybromo-1 bromodomain missense variants associated with clear cell renal cell carcinoma exhibit variable biochemical fitness, Karina Bursch, Medical College of Wisconsin

The tumor suppressor Polybromo-1 (PBRM1) modulates chromatin accessibility via six bromodomains that bind acetylated nuclear proteins. PBRM1 mutations exist in ~40% of clear cell renal cell carcinoma (ccRCC) cases, often ablating protein expression and tumor suppressor activity. However, missense mutations cluster in PBRM1 bromodomains and exist in ~15% of ccRCC cases, yielding full-length protein variants with unknown structural and functional characteristics. With >900 PBRM1 missense variants reported in the Catalogue of Missense Mutations in Cancer, mechanistic studies of every missense variant are intractable; *in silico* approaches may instead yield actionable assessments of PBRM1 missense variant impacts in ccRCC and other cancer types. We computationally predicted the impacts of ccRCC-associated missense variants in the second (BD2) and fourth bromodomains (BD4) of PBRM1 on

bromodomain stability and ligand binding at the levels of protein sequence (2D), structure (3D), and molecular dynamics (4D). These predictions were used to assign missense variants a pathogenicity score between 0-1; scores near 1 indicate missense variants predicted to decrease stability and ligand binding. We purified all reported ccRCC-associated PBRM1-BD2 and -BD4 missense variants to evaluate the concordance between predicted pathogenicity and experimental stability (differential scanning fluorimetry, circular dichroism), and ligand binding (AlphaScreen assays, electrophoretic mobility shift assays). 3D and combined 3D/4D scores correlated with measures of acetylated ligand binding, while 2D and total pathogenicity scores correlated with measures of protein thermal stability. Our results demonstrate that computational prediction of PBRM1 missense variant pathogenicity accelerates mechanistic understanding of missense variant impacts on protein stability and ligand binding, supporting future studies correlating specific missense variants with PBRM1 tumor-suppressive functions.

Comprehensive Independent Component Analysis Denoising Assistant (CICADA): A new simple and comprehensive fMRI denoising tool, Keith Dodd, University of Colorado

BACKGROUND: Manual independent component (IC) classification, a gold-standard functional magnetic resonance imaging (fMRI) denoising method, is time and training intensive. Automated methods, meanwhile, may denoise less effectively. To address this, we developed the Comprehensive Independent Component Analysis Denoising Assistant (CICADA) to better target all common noise sources and therefore hypothesized denoising metric improvements over current automated denoising methods. **METHODS:** CICADA was assessed in schizophrenia resting-state fMRI (N = 55) against FIX, ICA-AROMA, eight parameter (8p) regression, and manual IC denoising in a data subset (N = 20). Denoising efficacy was evaluated through gray matter connectivity correlations with motion (median framewise displacement [FD]) and distance dependence. Modularity quality and its correlation to motion evaluated network identifiability. Denoising effectiveness was further evaluated through noise profile correlations and IC classification accuracy. **RESULTS:** CICADA significantly reduced correlations between gray matter connectivity and both motion ($p < 0.001$ for all techniques) and distance dependence ($p < 0.001$ for 8p). 8p's moderate motion correlation ($r = 0.42$) suggests that the increased modularity quality is reflective of motion effects rather than neuronal activity. CICADA further consistently outperformed FIX, AROMA, and 8p in noise profile correlations and improved IC classification accuracy compared to AROMA and FIX ($p < 0.001$). **CONCLUSION:** CICADA appears to offer similar fMRI denoising effectiveness in resting-state fMRI to the manual IC denoising gold standard, with significant improvements compared to current automated methods.

Concurrent Session 2B

II. Cancer Biology (Elevation Ballroom)

Evidence of microsatellite-instability and high tumor mutational burden in HIV+/ART-experienced diffuse large B-cell lymphoma, Sophia Roush, University of North Carolina at Chapel Hill

Background: Diffuse large B-cell lymphoma (DLBCL) is highly associated with HIV and outcomes remain poor for many patients. With improved global access to antiretroviral therapy (ART), the proportion of DLBCLs arising in HIV+/ART-experienced (HIV+/ART-exp) people is increasing. Mutational profile, tumor mutational burden (TMB) and microsatellite-instability (MSI) are used as biomarkers in a variety of cancers, particularly for immunotherapy response, but have not been well studied in HIV+ DLBCL. **Methods:** We performed whole exome sequencing of n=24 DLBCL tumors and matched germline of patients enrolled in the Kamuzu Central Hospital Lymphoma Study (n=8 HIV-, n=12 HIV+/ART-exp, n=4 HIV+/ART-naïve). Pairwise Wilcoxon rank-sum test was used to compare TMB by HIV/ART status. “High” vs. “low” TMB was determined by median cut-off and difference in 5-year OS was assessed by log-rank test. For mutation enrichment analysis, pairwise Fisher’s exact test was used with FDR correction. MSI was predicted by the Microsatellite Analysis for Normal-Tumor InStability pipeline. Protein expression of MLH1, MSH6, MSH2, PMS2 was determined by immunohistochemistry (IHC). **Results:** Median TMB was higher in HIV+/ART-exp vs. HIV- DLBCL (1.6-fold, p=0.04), and TMB was positively prognostic among HIV+/ART-exp patients (20 vs. 10 mo. median survival, p=0.016), but not in the overall cohort. HIV+ DLBCLs were enriched for *ARID1B*, *CHD5*, *POLE*, *PTPRB*, *RELN*, and *RNF213* mutations, while HIV- DLBCLs were enriched for *NCOR2* and *CARD11* mutations. n=3 HIV+/ART-exp and n=1 HIV- DLBCLs were predicted to be MSI-positive, supported by mutations in DNA mismatch repair genes and protein loss by IHC. **Conclusion:** HIV+/ART-exp DLBCLs demonstrated increased TMB, and high TMB associated with improved survival in this group only. Our data suggest that MSI and defective DNA damage repair mechanisms play a role in HIV+/ART-exp DLBCL, and these patients may especially benefit from immune checkpoint inhibitor therapy.

Malignant mast cells are chemoresistant and associated with poor outcome in core binding factor-mutated pediatric acute myeloid leukemia, Denis Ohlstrom, Emory University/Georgia Institute of Technology

Background. Pediatric acute myeloid leukemia (pAML) is genetically heterogenous with malignancy driving translocations such as RUNX1-RUNX1T1 and CBFB-MYH11, collectively the core binding factor (CBF) pAMLs. Mutations inform pre-treatment risk, though 5-year relapse rates remain ~30% for standard risk (CBF) patients. Single-cell RNA sequencing (scRNAseq) enabled characterization of cellular diversity in pAML, but it is unclear which cells survive treatment and promote relapse. **Methods.** We analyzed 649,192 cells from 167 bone marrow (BM) biopsies of 98 patients (37 with sequential samples). PAML cells were identified by occupancy scoring, copy number variants, and immunophenotyping. Differentially expressed genes (DEG) were used for pathway analysis and deconvolution of the TARGET bulk RNAseq dataset. High-risk subtypes were defined by proportional increase at end-of-induction (EOI), enrichment in patients who experienced relapse, and worse overall survival (OS) in TARGET. **Results.** We identified 33 subtypes of pAML by cell surface gene expression. Surprisingly, one subtype expressed mast genes (e.g. *TPSB2* Log₂FC=4.92, p=1.0x10⁻²⁵⁰). Malignant mast cells (MMC) were specific to CBF pAML, with >5% MMCs in ~29% CBF patients vs 1% non-CBF. MMCs were chemoresistant (5.74-fold proportional increase at EOI vs diagnosis) and enriched in patients who experienced relapse vs sustained remission (2.38-fold). Patients enriched for MMCs in TARGET had worse OS (HR=2.24, p=6.7x10⁻⁴). DEGs included markers of immaturity (GATA2

$\text{Log}_2\text{FC}=2.63$, $p=1.0 \times 10^{-250}$) and immune interaction (CXCL8 $\text{Log}_2\text{FC}=2.53$, $p=1.0 \times 10^{-250}$). Pathway analysis similarly found enrichment for inflammation and immune cell chemotaxis. Conclusions. We report the first description of MMCs in CBF pAML. MMCs are chemoresistant, relapse-enriched, and associated with worse OS. MMCs express mixed mature and immature genes and immune interacting pathways. Future studies will dissect the spatial interaction of MMCs and immune cells in the BM.

Patient-derived glioblastoma organoids as real-time avatars to assess responses to clinical CAR-T cell therapy, Yusha Sun, University of Pennsylvania

Patient-derived tumor organoids have been increasingly leveraged for disease modeling and preclinical studies, but rarely in real-time to aid with the interpretation of patient treatment responses in the clinical setting. Whether tumor organoids can be applied to mirror clinical activity in patients and even to predict responses for personalized medicine remains uncertain. We recently demonstrated promising early efficacy signals in a first-in-human, phase 1 study of dual-targeting chimeric antigen receptor T cells (EGFR-IL13R α 2 CAR-T cells) in patients with recurrent glioblastoma (clinical trial identifier: NCT05168423; Bagley et al. Nature Medicine 2024). Determination of meaningful clinical efficacy, in particular for CAR-T cell therapy, may take months to manifest. Given this challenge, real-time analysis of patient-derived glioblastoma organoids (GBOs) could help to provide early assessment of clinical efficacy and guide patient management. Here we analyzed six sets of GBOs generated on the same timeline as the production of patients' CAR-T cell products. GBOs demonstrated the presence and retention of both targeted tumor-associated antigens EGFR and IL13R α 2 by immunohistochemistry. These GBOs were then treated with the same autologous CAR-T cell products received by patients in our phase 1 study in parallel with patient treatment. CAR-T cell treatment led to target antigen reduction, and we confirmed cytolysis of tumor cells in GBOs by cleaved caspase-3 immunostaining and by co-culture with the Axion Maestro Cellular Impedance platform. Importantly, the degree of cytolysis *ex vivo* significantly correlated with CAR-T cell engraftment detected in patients' cerebrospinal fluid (CSF) for these patients. Furthermore, cytotoxic activity of CAR-T cells as measured by TNF α , IL-2, and IFN γ release kinetics in GBOs mirrored cytokine levels in patient CSF samples over time. Our findings highlight a unique trial design and suggest GBOs as a valuable platform for real-time assessment of CAR-T cell bioactivity and for insights into immunotherapy efficacy.

Concurrent Session 3A

I. Neuroscience (Blue Spruce/Aspen)

SUR1-TRPM4 expression due to chronic seizures contributes to epileptogenesis, Mitchell Moyer, University of Maryland

Rationale: One third of epilepsy patients become resistant to current anti-seizure therapies. A promising novel target for reducing seizures in epilepsy is SUR1-TRPM4, a sodium channel minimally expressed in healthy brain that upregulates *de novo* after status epilepticus and other epilepsy-associated pathologies. SUR1-TRPM4 inhibition was shown to be therapeutically beneficial in numerous CNS pathologies in preclinical and clinical studies; however, this has not been explored in epilepsy. Here we investigated whether SUR1-TRPM4 upregulation increases chronic seizure susceptibility in epilepsy. **Methods:** SUR1-TRPM4 expression was assessed by immunohistochemistry in resected brain tissues electrographically sorted as normal or epileptic from epilepsy patients. To investigate the mechanism that SUR1-TRPM4 promotes seizures through neuron hyperexcitability, cortical cultures were recorded with microelectrode array, treated with low Mg^{2+} to induce hyperactivation, and co-treated with TRPM4 inhibitor CBA. To assess effects of SUR1-TRPM4 inhibition on chronic seizure susceptibility, a pentylenetetrazol (PTZ) kindling epilepsy model was used in mice. SUR1-TRPM4 was inhibited during PTZ kindling either pharmacologically by the FDA-approved SUR1 inhibitor glyburide or the TRPM4 inhibitor 9-phenanthrol, or genetically by constitutive or neuron-specific TRPM4 knock-out. **Results:** Compared to controls, SUR1-TRPM4 expression was increased within epileptic tissues resected from patients ($p < 0.05$, paired t-test). Inhibition of SUR1-TRPM4 *in vitro* reduced neuronal hyperactivity induced by low Mg^{2+} ($p < 0.01$, 2-way ANOVA). In all PTZ experiments, SUR1-TRPM4 inhibition significantly attenuated seizures ($p < 0.0001$, logistical regression). **Conclusions:** This study suggests SUR1-TRPM4 expression is upregulated in epilepsy and increases chronic seizure susceptibility through neuronal hyperexcitation. SUR1-TRPM4 may be a clinically relevant biomarker and therapeutic target to reduce seizures.

A tale of two cell types: calcitonin gene related peptide activates neurons and oligodendrocyte precursors cells in a sex- and region-dependent fashion, Rebecca Lorsung, University of Maryland

Calcitonin gene-related peptide (CGRP)-expressing neurons in the parabrachial nucleus (PB) densely project to several brain structures implicated in the emotional processing of stressful stimuli, including the extended amygdala and the insula cortex. We are testing the overarching hypothesis that PB CGRP release in these structures is causally involved in anxiety. Using *in vivo* fiber photometry of a peptide sensor, we found that PB CGRP-expressing neurons release large dense core vesicles, containing neuropeptides including CGRP, in the extended amygdala in response to an acute stressful stimulus – foot shock. Once CGRP is released, it can directly modulate brain activity via CGRP receptors on neurons, or indirectly via receptors on oligodendrocyte precursor cells (OPCs). Using a combination of whole-cell, voltage-clamp electrophysiology in live brain slices, as well as *in vitro* culturing of primary mixed brain culture, we discovered that CGRP exerts a sex- and region-dependent effect on these two cell types. In

the extended amygdala, CGRP preferentially promotes hyperexcitability in female neurons by potentiating neuronal responsiveness to excitation while simultaneously decreasing responsiveness to inhibition (disinhibition). Male extended amygdala neurons are partially protected due to a compensatory, CGRP-dependent increase in inhibitory tone. However, in the insula cortex, CGRP acts exclusively via CGRP receptors on OPCs; neither endogenous CGRP release from PB terminals, nor acute application of exogenous CGRP, alters insula neuronal responsiveness. These findings are supported by single cell RNAseq studies in both mice and humans, and RNAscope in mice, where we confirmed that only OPCs, not neurons, express CGRP receptors in the insula cortex. We are investigating whether CGRP in the extended amygdala plays a causal role in driving sex differences in anxiety and pain behaviors, and determining how CGRP-dependent activation of OPCs alters neuronal activity.

Early life injury alters spinal astrocyte development, Judy Yoo, University of Cincinnati

9% of children born in the United States are admitted to the neonatal intensive care unit where many experience invasive surgical procedures as part of their medical care. Exposure to invasive procedures in early life increases the excitability of the spinal nociceptive network and contributes to 'priming' of the nociceptive pathway, causing increased pain perception in response to noxious stimuli later in life. Astrocytes orchestrate synapse development and function; however, nothing is currently known about how developing spinal astrocytes respond to early life injury. Thus, we sought to elucidate the effects of neonatal surgical injury on the structure and transcriptional profile of astrocytes in the developing dorsal horn. At postnatal day 3 (P3), mice expressing tdTomato in astrocytes received either a unilateral incision of the hindpaw skin and underlying muscle or anesthesia only as controls. We then imaged astrocytes in the ipsilateral dorsal horn at P4, P10, and P24 using confocal microscopy in cleared tissue for accurate 3D reconstruction. Using quantitative morphological analysis, we found that astrocytes in the P3 incision group exhibit increased cell size and complexity at P4, while their size and complexity were reduced at both P10 and P24 when compared to naïve controls. Next, we used mice with green fluorescent protein (GFP)-expressing astrocyte nuclei and performed bulk RNA sequencing on sorted GFP+ nuclei to identify differentially expressed genes (DEGs) in astrocytes between incised mice and naïve controls. Our transcriptomic analysis revealed 76 DEGs between naïve and incision groups at P4, 2 DEGs at P10, and 8 DEGs at P24. Among these DEGs, we found that genes coding for matricellular proteins such as *Thbs1* and *Efemp1* and cytoskeletal genes such as *Acta1* and *Acta2* are upregulated at P4 following incision. Our data show, for the first time, that early life injury evokes morphological and transcriptional changes in developing spinal astrocytes.

Concurrent Session 3B

II. Molecular Biology and Pharmacology (Elevation)

RyR2 mediated calcium dysregulation increases glyoxalase I in the aging brain, Elizabeth Woo, Yale School of Medicine

The greatest risk factor for sporadic Alzheimer's disease is aging. With age, there is loss of regulation of cAMP-PKA-calcium signaling, which is critical for supporting working memory, leading to chronically elevated cytosolic calcium. Specifically, phosphorylation of ryanodine receptor 2 (RyR2) at S2808 causes calcium to "leak" from the smooth endoplasmic reticulum, occurring in Alzheimer's disease, chronic stress models, and long COVID-19. However, the potential compensatory mechanisms utilized by neurons in response to calcium dysregulation are not well understood. Our study utilized a genetic mouse model, S2808D-RyR2 (S2808D), causing a constitutive calcium leak through a knock-in mutation, and the aging rhesus macaque model. We performed mass spectrometry on synaptosomes from the frontal cortex (FC) and hippocampus (HP) of 3-month-old mice, followed by Ingenuity Pathway Analysis. Among the significantly differentially expressed proteins, glyoxalase 1 (Glo1) was 2.17 times higher ($p=0.0014$) and 2.22 times higher ($p=0.011$) in the S2808D FC and HP synaptosomes, respectively. All findings were validated using immunoblotting techniques. S2808D synaptosomes also had significantly increased Glo1 activity in the FC ($p=0.0015$) and the HP ($p=0.0146$). Glo1 is a critical enzyme responsible for removing glycative stress, and it has been implicated in diabetes, neurodegeneration, and psychiatric disorders. The significant increase in Glo1 expression and activity was found in adult S2808D FC and HP synaptosomes. In aged rhesus macaques, we also found significantly increased Glo1 expression in the prefrontal cortex (PFC) ($p=0.0381$), indicating this restorative mechanism also occurs in primates. Pairing MLIF and immuno-electron microscopy, we quantified and characterized the localization of Glo1 for the first time in macaque PFC. Taken together, these findings suggest that elevated Glo1 expression may provide resilience in response to dysregulated calcium levels in neurons.

PDGFR α signaling regulates Srsf3 transcript binding to affect PI3K signaling and endosomal trafficking Thomas Forman, University of Colorado

Signaling through the platelet-derived growth factor receptor alpha (PDGFR α) plays a critical role in craniofacial development, as mutations in *PDGFRA* are associated with cleft lip/palate in humans and *Pdgfra* mutant mouse models display varying degrees of facial clefting. Phosphatidylinositol 3-kinase (PI3K)/Akt is the primary effector of PDGFR α signaling during skeletal development in the mouse. We previously demonstrated that Akt phosphorylates the RNA-binding protein serine/arginine-rich splicing factor 3 (Srsf3) downstream of PI3K-mediated PDGFR α signaling in mouse embryonic palatal mesenchyme (MEPM) cells, leading to its nuclear translocation. We further showed that ablation of *Srsf3* in the murine neural crest lineage results in severe midline facial clefting, due to defects in proliferation and survival of cranial neural crest cells, and over a thousand differential alternative RNA splicing events. Here, we demonstrate via enhanced crosslinking and immunoprecipitation (eCLIP)-seq analysis of MEPM cells that PDGF-AA stimulation leads to preferential binding of Srsf3 to exons. Further, an unbiased motif enrichment analysis of Srsf3 binding sites revealed a loss of binding to canonical Srsf3 CA-rich motifs in stimulated samples, which could be due to changes in ribonucleoprotein composition

or loss of RNA-binding due to electrostatic repulsion. Through the analysis of complementary RNA-seq data, we show that the subset of transcripts that are bound by Srsf3 and undergo alternative splicing upon PDGFR α signaling commonly encode regulators of Wnt signaling, a pathway known to be critical for mammalian craniofacial development. Taken together, these findings provide considerable insight into the mechanisms underlying gene expression regulation during mammalian craniofacial development.

The non-coding cause of congenital heart defect: abnormal RNA splicing with multiple novel isoforms as a mechanism for heterotaxy, John Wells, Indiana University

Heterotaxy is a disorder characterized by severe congenital heart defects (CHDs) and abnormal left-right patterning in other thoracic or abdominal organs. Clinical and research-based genetic testing has previously focused on evaluation of coding variants to identify causes of CHDs, leaving non-coding causes of CHDs largely unknown. Variants in the transcription factor Zinc finger of the cerebellum 3 (*ZIC3*) cause X-linked heterotaxy. We identified an X-linked heterotaxy pedigree without a coding variant in *ZIC3*. Whole genome sequencing revealed a novel, deep intronic variant (*ZIC3* c.1224+3286A>G) predicted to alter RNA splicing. An *in vitro* minigene splicing assay confirmed the variant acts as a cryptic splice acceptor. CRISPR/Cas9 served to introduce the *ZIC3* c.1224+3286A>G variant into human embryonic stem cells demonstrating pseudoexon inclusion caused by the variant. Surprisingly, Sanger sequencing of the resulting *ZIC3* c.1224+3286A>G amplicons revealed several novel isoforms, many of which by-pass the normal coding sequence of the third exon of *ZIC3*, causing a disruption of a DNA binding domain and a nuclear localization signal. Short- and long-read mRNA sequencing confirmed these initial results and identified additional novel splicing patterns. Assessment of four novel isoforms determined an abnormal function *in vitro* and *in vivo* while treatment with a splice-blocking morpholino partially rescued *ZIC3*. These results are the first reported instance of pseudoexon inclusion in heterotaxy and provide functional validation of non-coding disease causation. Our results suggest the importance of non-coding variants in heterotaxy and the need for improved methods to identify and classify non-coding variation that may contribute to CHDs.

Poster Sessions

Poster Session I

Sponsored by FAER

1. Maya Anand

University of Rochester

Discussion Prioritization Tool Eliciting Preferences of Older Adults with Breast Cancer Considering Adjuvant Chemotherapy: A Pilot Usability Study

2. Adriana Baker

South Texas MSTP at University of Texas Health San Antonio

ERX-315: A Potential Treatment Option for Hepatocellular Carcinoma (HCC)

3. Emily Balczewski

University of Michigan MSTP

Computable phenotyping of antiphospholipid syndrome with electronic health record data

4. Carl Bannerman

Perelman School of Medicine

The Role of Interferon Epsilon in Human Neurons

5. Riley Behan-Bush

University of Iowa

Polychlorinated biphenyls induce macrophage immunometabolic plasticity

6. John Bennett

University of Rochester

Chemoenzymatic Synthesis and Anticancer Activity of a New Class of Skeletally Diverse Natural Product-like Compounds

7. Caylin Billingsley

Indiana University School of Medicine

Cellular Growth Mindset: HPV 16E6 and NFX1-123 Partnership Amplifies Proliferation of Long-Term Keratinocyte Cultures

8. Stephanie Busch

Zucker School of Medicine at Hofstra/Northwell

Isolation, in-vitro expansion, and characterization of menstrual effluent-derived uterine natural killer cells

9. Justin Campbell

University of Utah School of Medicine

DIRECT ELECTRICAL STIMULATION OF THE BASOLATERAL AMYGDALA MODULATES OSCILLATORY DYNAMICS IN THE HIPPOCAMPUS

10. Nicholas Chargo

Michigan State University College of Osteopathic Medicine DO/PhD Program

Dynamic effects of dietary prune on the gut-bone axis in healthy female mice

11. Steve Cho

University of Utah MSTP

Directed evolution of a nascent immune function

12. Aidan Crowley

University of Pennsylvania Perelman School of Medicine
Performance of Physician Groups and Hospitals in Bundled Payments for Care Improvement Advanced for Lower Extremity Joint Replacement

13. Paarth Dodhiawala

University of Minnesota Medical School
KRAS inhibition enhances NK cell-mediated tumor killing via potential ULBP an IL-15 mediated mechanism in pancreatic cancer

14. Nada Elsayed

Vanderbilt University
THE CELLULAR LANDSCAPE OF CORTICAL DEVELOPMENT IN TUBEROUS SCLEROSIS COMPLEX

15. Brianna Evans

Pennsylvania State College of Medicine
Acute treatment with the glucagon-like peptide-1 receptor agonist, liraglutide, reduces cue-induced fentanyl seeking and associated c-Fos and tyrosine hydroxylase activation patterns in brain in rats

16. Ilaria Ferrari

University of Louisville
Exercise Prior to Myocardial Infarction has Minimal Impact on Ventricular Remodeling

17. Sarah Florig

University of Nebraska Medical Center
The Impact of Telehealth on Clinician Ordering Behaviors and Test Completion Rates

18. Arlene Garcia

Vanderbilt University
Elucidating mechanisms of mitochondrial DNA copy number decline in aging

19. Elsie Gonzalez-Hurtado

Yale University School of Medicine
Nerve-associated macrophages control adipose homeostasis across lifespan and restrain age-induced inflammation

20. Freedom Green

South Texas MSTP at University of Texas Health San Antonio
Leading the Pack: Strategies for Supporting Service Dog Handlers and Individuals with Disabilities and Chronic Illnesses in STEM

21. Candace Grisham

Vanderbilt University
Role of Fibroblast Activation Protein in Cancer Therapeutic Efficacy

22. Cristhian Gutierrez Huerta

Medical College of Wisconsin MSTP
Mitochondrial fission in the microvascular endothelium leads to microvascular dysfunction in humans

23. Alexandra Hommer

Cornell University Combined DVM/PhD

The role of subcellular protein methionine oxidation and methionine sulfoxide reductase (MSR) activities in aging

24. Fengling Hu

University of Pennsylvania School of Medicine

Automated Segmentation of Multiple Sclerosis Lesions, Paramagnetic Rims, and Central Vein Sign on MRI

25. Miranda Jankovic

UT Southwestern Medical Center

The Foxp2 Influence on the Developing Auditory Pathway

26. Claudia Rose Keating

Loyola University Chicago

Characterizing the role of connective tissue growth factor in hepatocellular carcinoma

27. Thomas Kelly

Medical College of Wisconsin

Selective deletion of mTOR in midbrain dopamine neurons impairs dopamine transmission and induces attention deficiency, hyperactivity and impulsivity

28. Ruchita Kothari

Johns Hopkins MSTP

Investigating the Role of a Mast Cell-Specific Receptor in Mediating Stroke Associated Inflammation

29. Curtis Kuo

University of Michigan MSTP

Loss of Myonuclear Type IIb and IIx Specificity in Spinal and Bulbar Muscular Atrophy

30. Jessica Lam

University of Pennsylvania School of Medicine

YY1 controlled regulatory connectivity and transcription ?are influenced by the cell cycle

31. Kaylin Langer

University of Colorado School of Medicine

Oligodendrocyte response and myelin composition following neonatal hypoxic ischemic brain injury

32. Catherine Li

University of North Carolina at Chapel Hill MD-PhD Program

Per-protocol effects of dolutegravir-based therapy compared to standard of care in children living with HIV in the ODYSSEY trial

33. Mark Libowitz

University of Utah MSTP

Underlying Effects of Intracranial Stimulation on Insula Functional Connectivity

34. Samuel Liburd

Yale University

Vascular phenotype as predictive markers of CAR-T cell therapy response in patients with diffuse large B-cell lymphoma

35. Ryan Mack

Loyola University Chicago

RESOLUTION OF COMMON MYELOID PROGENITOR HETEROGENEITY AND IDENTIFICATION OF A NOVEL DIFFERENTIATION PATHWAY

36. Charles Marcucci IV

University of Virginia MSTP

Deciphering the role of tenascin-n in peripheral spinal motor nerve development

37. Michael Meadow

MSTP at University of Rochester School of Medicine & Dentistry

Proteome birthdating reveals age-selectivity of protein ubiquitination

38. Bani Medegan Fagla

University of Illinois College of Medicine MSTP Program

APOE4 genotype increases susceptibility to fetal growth restriction in the context of preeclampsia in a human cohort and in a mouse model of preeclampsia

39. Amanda Paskavitz

Wayne State University School of Medicine

Second-generation sperm epigenetic clock predicts male fertility age and time-to-pregnancy in general population

40. Gavin Piester

University of Rochester School of Medicine & Dentistry

THE SIGMA-1 RECEPTOR DRIVES ASTROCYTE PATHOGENIC ACTIVITIES IN PARKINSON'S DISEASE

41. John Quinlan

University of Maryland, College Park

Photodynamic Therapy in Glioblastoma Multiforme using Self-Assembled Verteporfin Nanoparticles

42. Sarah Reed

Vanderbilt University MSTP

Clonal hematopoiesis of indeterminate potential alters tumor microenvironment and clinical outcomes in solid tumors

43. Kishan Sangani

University of Chicago

Gut Check: Tet2 and Th2 Responses in the Intestine

44. Luke Schroeder

University of Louisville

Advancement of a 3D human U87 glioblastoma spheroid suspension model to augment traditional 2D tumor-derived small extracellular vesicle investigations

45. Kallen Schwark

University of Michigan

Optimizing PDGFRA Inhibition Therapy for Pediatric High-Grade Glioma

46. Karishma Shah

University of Utah MSTP

When We Become Our Pain

47. William Sheeran

University of Colorado

Ventral hippocampal contributions to lasting social memory in prairie voles.

48. Samantha Stone

University of Utah MSTP

Mechanistic Interactions Between Depression, Selective Serotonin Reuptake Inhibitors (SSRIs), And Cancer Patient Survival

49. Rhea Sullivan

Penn State College of Medicine

Opioid-responsive microRNAs in brain stem cell fates: implications for neonatal opioid withdrawal syndrome infants

50. Maya Talukdar

Harvard/MIT MD-PhD Program

Genes of the fatty acid oxidation pathway are upregulated in the female as compared to male human heart

51. Kaylee Tutrow

Indiana University School of Medicine

A human induced pluripotent stem cell model of Alzheimer's Disease-associated fractalkine receptor polymorphism to assess AD-related microglial dysfunction

52. Mayuri Viswanathan

University of Chicago MSTP

Gamma constant usage tunes $\gamma\delta$ T cell receptor sensitivity, thymic programming, and peripheral function

53. Qi Wang

University of Iowa Carver College of Medicine

Dynamic Mechanical Loading of Photopolymerized Hydrogels for Investigating the Interplay of Mechanical and Biochemical Signals in Pulmonary Fibrosis

54. Nathaniel Wright

University of Chicago

BRWD1 establishes epigenetic states for germinal center initiation, maintenance, and function

55. Crystal Young

University of Michigan MSTP

Hypothyroidism with Defective Thyroglobulin Co-opts the Machinery of Thyroid Hormone Synthesis to Drive Thyroid Cell Death

56. Ashley Zachery-Savella

University of Utah MSTP

Using Seizure Onset and Offset Patterns to Predict Drug Response in an Animal Model of Temporal Lobe Epilepsy

57. Walter Zhao

Case Western Reserve University

Automatic Risk Priors for Prediction of Glioblastoma Peritumoral Infiltration Using MR Fingerprinting and Multiparametric MRI

Poster Session II

Sponsored by Department of Pathology University of Colorado Anschutz

- 1. Joshua Baker**
Michigan State University College of Osteopathic Medicine
Insights of Contemporary Diffusion MRI Modeling Techniques in Mild Traumatic Brain Injury: A TRACK-TBI Study
- 2. Eric Barrientos**
University of Colorado
Shear-Dependent Volume Analysis of Platelet Aggregates under Pharmacological Inhibition in Microfluidic Devices
- 3. Ben Battistone**
University of Utah
T cell miR-155 Regulates mRNA Vaccine Immunotherapy Against Acute Myeloid Leukemia
- 4. Riley Bergman**
Vanderbilt University School of Medicine
The role and tolerability of SF3B1 hotspot mutations in breast cancer
- 5. Daniel Caballero**
Perot Family Scholars MSTP at UT Southwestern
The yeast Mkt1/Pbp1 complex promotes adaptive responses to respiratory growth
- 6. Hannah Carter**
University of Michigan
Dendritic Cell - Fibroblast Crosstalk Via Toll-like Receptor 9 and Aryl Hydrocarbon Receptor Signaling Drives Lung Fibrogenesis
- 7. Thomas Delgado**
University of Rochester
Depletion of Astrocytic TG2 Enhances the Ability of Astrocytes to Metabolically Support Neurons through Lipid Metabolism
- 8. Julie Disharoon**
Medical University of South Carolina MSTP
The role and regulation of BRD4 in the DNA Damage Response
- 9. Caroline Doherty**
Mayo Clinic
In Vivo Selection of anti-GBM DNA aptamers in an orthotopic patient derived xenograft model
- 10. Jacob Edmondson**
University of Arkansas for Medical Sciences
ATF6 activation in melanoma promotes anti-tumor immunity and improves ICB therapy response
- 11. Eric Engelbrecht**
University of Louisville School of Medicine MD/PhD Program

Impact of Human Immunoglobulin Genetic Variants on the Expressed Antibody Repertoire

12. Alan Finkelstein

University of Rochester

Integrated Microvascular and Microstructural Characterization through Intravoxel Incoherent Motion Tensor Imaging

13. Kevin Fundora

Penn State College of Medicine

Targeting VPS4 as a novel anticancer therapy

14. Kerrie Greene

Yale University

Intranasal Delivery of CD4 T Cell Peptides for Enhanced Therapeutic Access to the CNS

15. Julian Grosskopf

Medical College of Wisconsin

Modeling protein conformational ensembles using experimental DEER spectroscopy data

16. Samson Hennessy-Strahs

Texas A&M School of Medicine

Conditional Deletion of mTOR/TRAF6 Reduces Alveolar Macrophages and Leads to Alveolar Proteinosis

17. Laurel Hiatt

University of Utah

STRategies for STRs' STRangeness with STRchive, a dynamic resource for short tandem repeat disease loci in humans

18. Reuben Hogan

University of California, San Francisco

Comparâtes Analysis of Glycoproteomic Software Tools Using a Tailored Glycan Database

19. Anna Hudson

University of Miami Miller School of Medicine

Overcoming the Immunosuppressive Glioblastoma Microenvironment by targeting HERV-K

20. Katherine Irwin

Johns Hopkins MSTP

A fluid biomarker reveals loss of TDP-43 splicing repression in presymptomatic ALS-FTD

21. Amanda Jiang

University of Utah

Unraveling MC1R Gene Variants in Lentigo Maligna Melanoma in the Utah Population

22. Austin Jolly

University of Colorado

Remodeling of the vascular wall is regulated by the chromatin remodeler Brg1

23. Jakub Kaczmarzyk

Stony Brook University

Towards transparent and trustworthy AI in precision oncology and computational pathology

24. Shanawaj (Roy) Khair

University of Colorado School of Medicine

ROLE OF ANTIMICROBIAL PEPTIDES IN POST-BURN PULMONARY INFLAMMATION

25. Katie Kruk

University of Maryland, Baltimore

A Mechanism by Which Estradiol (E2) Regulates Adenosinergic Signaling in the Median Preoptic Nucleus (MnPO)

26. Michael Lee

University of Texas Southwestern

LINE-1 retrotransposon transcription mediates oncogenic chromatin interaction hubs

27. Siying Lin

Medical College of Wisconsin

Satb1 regulates the stemness of antigen-specific CD8 T+ cells

28. Matthew Loberg

Vanderbilt University

Multi-omic analysis identifies fibroblast subpopulations associated with aggressive thyroid cancer behavior

29. Alexander Lu

Texas A&M College of Medicine

O-GlcNAcylation Regulates Angiogenic Transdifferentiation Through HIRA Dependent H3.3 Deposition

30. Adarsha Malla

University of Maryland, Baltimore

Adjuvant transcranial microbubble-enhanced focused ultrasound treatment of infiltrating gliomas in humans and rodents

31. Anna Maximova

University of Maryland Baltimore

Investigating the impact of a unique peri-hippocampal mast cell population in postnatal rat neurodevelopment

32. Jennifer Messina

SUNY Upstate Medical University

Battling Hyperinflammation and Pyroptosis in Sepsis Using Bioactive Nanoparticles

33. Raymond Ng

University of Pennsylvania School of Medicine

The role of tumor transcriptional variability in evading acute CD8+ T cell exposure in melanoma

34. Fae Nova

Pennsylvania State University College of Medicine

A multimodal 3D developmental mouse brain common coordinate framework for cell census mapping

35. Ian Outhwaite

SUNY Stony Brook University

Kinase Inhibitor Combinations Maximize Selectivity and Enable Rational Multitargeting

36. Julia Phan

University of Texas Southwestern

Retrotransposons are Co-opted by Hematopoietic Stem Cells During Pregnancy to Increase Erythropoiesis

37. Randall Rainwater

University of Arkansas School for Medical Sciences

The Novel Role of DNA-PKcs in LAT-mediated T cell Signaling

38. Roshan Ravishankar

University of Texas Southwestern Medical Center

Automated Detection of Rho GTPase Signaling Microdomains

39. Melissa Requist

University of Utah MSTP

Proposed Mechanisms of Osseous Morphology Differences in Charcot Marie Tooth Disease

40. Juan Santiago Moreno

University of Colorado Anschutz Medical Program

THE DISTRIBUTION OF CHROMATIC TUNING IN THE MOUSE EARLY VISUAL SYSTEM

41. Vivien Sauer

University of Cincinnati College of Medicine MSTP

Investigating the role of SMAD6 in the development of tracheoesophageal birth defects

42. Ankita Saxena

SUNY UPSTATE MEDICAL UNIVERSITY

Application of Graph Neural Networks in Disease Risk Predictions: Augmenting GWAS with Novel Insights into Biological Networks

43. Justin Schumacher

University of Maryland School of Medicine

Clinical multimodal elastography for measuring spatial-varying mechanical properties of the crystalline lens.

44. Kayla Schwartz

University of Miami MSTP

Pancreatic cancers suppress fibroinflammatory signals to support metastatic outgrowth

45. Manlin Shao

University of Arizona College of Medicine-Tucson

Orthotopic and Immunocompetent Mouse Models for Oral Squamous Cell Carcinoma

46. Kelli Sommers

Medical College of Wisconsin

Immune-mediated microtubule destabilization induces axonal transport defects in a mouse model of optic neuritis

47. Jonathan Sussman

University of Pennsylvania School of Medicine

A longitudinal single-cell and spatial multiomic atlas of pediatric high-grade glioma

48. Josephine Trichka

Case Western Reserve University MSTP

CHMP5 restrains the tissue-intrinsic interferon response in skeletal muscle

49. Jude Tunyi

The Ohio State MSTP

Computational and Biochemical Characterization of Amino Acid Transporter

Protein-protein Interactions

50. Amanda Velez

University of North Carolina at Chapel Hill

Staphylococcus aureus pathogenesis within the polymicrobial diabetic chronic wound

51. Madison Wahlen

University of Iowa

Identifying Adaptations to Preserve Core Functions of an Evidence-Based

Intervention to Improve Cancer Care Quality in Rural Hospitals

52. Jenna Weber

University of Utah

Paternal Age Effect Mutations: Selfish Drivers of Germline Stem Cell Clonality?

53. Clare Wieland

University of Michigan

Neuron specific regulation of repeat associated non-AUG translation in age related dementias

54. John Yap

Loyola University Chicago, Stritch School of Medicine

Unraveling the role of PDK4 in sex dependent outcomes during sepsis induced cardiomyopathy

55. Grace Yu

Mayo Clinic MSTP

Effect of Senescent Melanocytes on Epidermal Skin Aging

56. Deanne Yugawa

Yale MD-PhD Program

ERK potentiates its own activity in pancreatic cancer via alterations in RNA splicing

57. Victor Zhang

University of Rochester

Type III collagen recreates the fibrotic myofibroblast microenvironment for studying peritendinous scarring in a human Tendon-on-a-Chip (hToC)

58. Sherry Zhou

Mayo Clinic M.D.-Ph.D. Program

Investigating post-translational protein acylation in SDH-loss pheochromocytoma and paraganglioma

2024 Travel Awardees

| Travel Awardees | |
|------------------------------------------------------------------|--------------------------------------------------------------|
| Brianna Evans , Pennsylvania State University | Marissa Chou , University of California San Francisco |
| Claudia Rose Keating , Loyola University Chicago | Caylin Billingsley , Indiana University |
| Kaylee Tutrow , Indiana University | Ashley Zachery-Savella , University of Utah |
| Siyang Lin , Medical College of Wisconsin | Jacob Edmonson , University of Arkansas |
| Cristhian Gutierrez Huerta , Medical College of Wisconsin | Jonathan Sussman , University of Pennsylvania |

Saturday Morning Activities

Breckenridge is an ideal location for exploring the Rocky Mountains! Several activities will be planned and organized by University of Colorado MSTP students. While these activities are subject to change due to inclement weather, they have usually happened as planned in years past.

Hiking

For all hikes, please bring the following: a pair of supportive shoes or boots, sunscreen, water, snack food, loose-fitting clothes, a warm layer, and a rain/wind-resistant layer.

Easy Hike – Minne Mine (2.9 mi 300 ft elev. gain.): Just a five-minute drive from Breckenridge, you and your friends will be positioned to explore the easy trails and explore old mining ruins. The hike from B&B to Reiling Dredge to Minnie Mine near Breckenridge offers a scenic and historical experience. Starting at the B&B Trailhead, the trail takes you through lush forests and past the historic Reiling Dredge, an old mining dredge from the early 1900s. The hike continues to Minnie Mine, where you can explore remnants of mining activity while enjoying picturesque views of the surrounding mountains. Breckenridge offers a shuttle service to trail head leaving every 30 min.

Meet at the Registration Table at 9:00am.

Intermediate Hike - Mayflower Lake (5.5 mi 1000 ft elev. gain.) The hike to Mayflower Lake near Breckenridge offers stunning alpine scenery, leading you past cascading waterfalls, historic mining ruins, and lush wildflower meadows, culminating in breathtaking views of the pristine high-altitude lakes.

Meet at the Registration Table at 8:00am.

Volleyball

There's nothing like a crazy game of volleyball to have a good time! We will set up a volleyball net at Carter Park and Pavilion about a 15-minute walk from the Conference Center. If you want to partake in some fun rallies, come join the game!

Meet at the Registration Desk at 9:15am.

Day Trip to Breckenridge

Downtown Breckenridge offers a charming blend of historic and modern attractions, with a vibrant Main Street lined with unique shops, art galleries, and a variety of food options. Visitors can explore cultural sites like the Barney Ford House Museum, which showcase the town's rich mining history. The downtown area features an array of specialty shops, including boutiques, souvenir stores, and artisan craft shops, providing a delightful shopping experience for all visitors.

Meet at the Registration Desk at 9:30am

Breakfast and Board Games

In case of inclement weather, or for those who prefer a more leisurely Sunday morning indoors, we will have a room reserved where students can enjoy a variety of traditional games such as Chess and GO as well as modern classics like Catan and Carcassonne. Players of all experience levels are welcome!

Meet in the Evergreen Ballroom at 9:30 am

**** All activities are weather dependent, non-refundable, non-transferrable, and we do not offer any refunds regardless of activity closures. Additionally, Activity Day Passes are day specific and we do not offer refunds on partially used day passes.**

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National Student Conference
July 12th - 14th
Breckenridge, CO

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Student Participants

| First Name | Last Name | University Program |
|------------|-----------------|------------------------------------------------------------|
| Mauricio | Alvarez | University of Colorado MSTP |
| Maya | Anand | University of Rochester |
| Adriana | Baker | South Texas MSTP at University of Texas Health San Antonio |
| Joshua | Baker | Michigan State University |
| Emily | Balczewski | University of Michigan MSTP |
| Amandip | Bangar | University of Colorado MSTP |
| Carl | Bannerman | Perelman School of Medicine |
| Eric | Barrientos | University of Colorado MSTP |
| Ben | Battistone | University of Utah |
| Hannah | Beatty | University of Colorado MSTP |
| Riley | Behan-Bush | University of Iowa |
| David | Beltran-Cardona | University of Colorado MSTP |
| John | Bennett | University of Rochester |
| Riley | Bergman | Vanderbilt University School of Medicine |
| Jessica | Beynor | University of Colorado MSTP |
| Caylin | Billingsley | Indiana University School of Medicine |
| Karina | Bursch | Medical College of Wisconsin |
| Stephanie | Busch | Zucker School of Medicine at Hofstra/Northwell |
| Daniel | Caballero | Perot Family Scholars MSTP at UT Southwestern |
| Alex | Camai | University of Colorado MSTP |
| Justin | Campbell | University of Utah School of Medicine |
| Hannah | Carter | University of Michigan |
| Sofia | Celli | University of Colorado MSTP |
| Nicholas | Chargo | Michigan State University College of Osteopathic Medicine |
| Michelle | Cho | University of Colorado MSTP Denver MSTP |
| Steve | Cho | University of Utah MSTP |
| Marissa | Chou | University of California, San Francisco |
| Rachel | Cohen | University of Colorado MSTP |
| Jacob | Connolly | University of South Florida |
| Aidan | Crowley | University of Pennsylvania Perelman School of Medicine |
| Thomas | Delgado | University of Rochester |
| Julie | Disharoon | Medical University of South Carolina MSTP |
| Keith | Dodd | University of Colorado MSTP |
| Paarth | Dodhiawala | University of Minnesota Medical School |
| Caroline | Doherty | Mayo Clinic |
| Jacob | Edmondson | University of Arkansas for Medical Sciences |
| Selin | Ekici | University of Colorado MSTP |

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|-----------------|------------------|------------------------------------------------------------|
| Nada | Elsayed | Vanderbilt University |
| Eric | Engelbrecht | University of Louisville School of Medicine MD/PhD Program |
| Elena | Esch | University of Colorado MSTP |
| Sofia | Esquibies | University of Colorado MSTP |
| Brianna | Evans | Pennsylvania State College of Medicine |
| Ilaria | Ferrari | University of Louisville |
| Alan | Finkelstein | University of Rochester |
| Erin | Fish | University of Colorado MSTP |
| Ira | Fleming | University of Colorado MSTP |
| Ian | Fleming | University of Colorado MSTP |
| Sarah | Florig | University of Nebraska Medical Center |
| Thomas | Forman | University of Colorado MSTP |
| North | Foulon | University of Colorado MSTP |
| Kevin | Fundora | Penn State College of Medicine |
| Allison | Galante | Renaissance School of Medicine at Stony Brook University |
| Arlene | Garcia | Vanderbilt University |
| Stefano | Ginocchio | University of Colorado MSTP |
| Volha | Golubeva | Ohio State University School of Medicine |
| Elsie | Gonzalez-Hurtado | Yale University School of Medicine |
| Freedom | Green | South Texas MSTP at University of Texas Health San Antonio |
| Kerrie | Greene | Yale University |
| Candace | Grisham | Vanderbilt University |
| Julian | Grosskopf | Medical College of Wisconsin MSTP |
| Cristhian | Gutierrez Huerta | Medical College of Wisconsin MSTP |
| Samson | Hennessy-Strahs | Texas A&M School of Medicine |
| Laurel | Hiatt | University of Utah |
| Brandon | Hilliard | University of Colorado MSTP |
| Reuben | Hogan | University of California, San Francisco |
| Alexandra | Hommer | Cornell University Combined DVM/PhD |
| Shannon | Hu | Renaissance School of Medicine at Stony Brook University |
| Fengling | Hu | University of Pennsylvania School of Medicine |
| Isabelle | Hua | University of Colorado MSTP |
| Anna | Hudson | University of Miami Miller School of Medicine |
| Katherine | Irwin | Johns Hopkins MSTP |
| Miranda | Jankovic | UT Southwestern Medical Center |
| Amanda | Jiang | University of Utah |
| Austin | Jolly | University of Colorado MSTP |
| Jakub | Kaczmarzyk | Stony Brook University |
| Raisa | Karim | Renaissance School of Medicine at Stony Brook University |
| Sritejasvinithi | Karimikonda | Medical College of Wisconsin |

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|----------------|---------------|----------------------------------------------------------------|
| Claudia Rose | Keating | Loyola University Chicago |
| Thomas | Kelly | Medical College of Wisconsin |
| Shanawaj (Roy) | Khair | University of Colorado MSTP |
| Ward | Kirschbaum | University of Colorado MSTP |
| Giona | Kleinberg | University of Colorado MSTP |
| Haley | Klimaszewski | The Ohio State University MSTP |
| Ruchita | Kothari | Johns Hopkins MSTP |
| Katie | Kruk | University of Maryland, Baltimore |
| Curtis | Kuo | University of Michigan MSTP |
| Jessica | Lam | University of Pennsylvania School of Medicine |
| Kaylin | Langer | University of Colorado MSTP |
| Michael | Lee | University of Texas Southwestern |
| Emily | Leventhal | Icahn School of Medicine at Mount Sinai |
| Steven | Lewis | Stony Brook University |
| Catherine | Li | University of North Carolina at Chapel Hill MD-PhD Program |
| Mark | Libowitz | University of Utah MSTP |
| Samuel | Liburd | Yale University |
| Shujian | Lin | University of Colorado MSTP |
| Siyang | Lin | Medical College of Wisconsin |
| Matthew | Loberg | Vanderbilt University |
| Vincent | Lok | University of South Florida Morsani College of Medicine |
| Rebecca | Lorsung | University of Maryland School of Medicine |
| Alexander | Lu | Texas A&M College of Medicine |
| Ryan | Mack | Loyola University Chicago |
| Adarsha | Malla | University of Maryland, Baltimore |
| Charles | Marcucci IV | University of Virginia MSTP |
| Matthew | Mardo | University of Colorado MSTP |
| Anna | Maximova | University of Maryland Baltimore |
| Nicholas | McQuillan | University of Colorado MSTP |
| Michael | Meadow | MSTP at University of Rochester School of Medicine & Dentistry |
| Bani | Medegan Fagla | University of Illinois College of Medicine MSTP |
| Jennifer | Messina | SUNY Upstate Medical University |
| Daniel | Moskop | University of Colorado MSTP |
| Mitchell | Moyer | University of Maryland |
| Rahul | Nachnani | Pennsylvania State University College of Medicine |
| Raymond | Ng | University of Pennsylvania School of Medicine |
| Fae | Nova | Pennsylvania State University College of Medicine |
| Younhye | Ock | University of Colorado MSTP |
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| Duby | Okonkwo | Vanderbilt School of Medicine MSTP |

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| Margo | Orlen | University of Pennsylvania School of Medicine |
| Ian | Outhwaite | SUNY Stony Brook University |
| Amanda | Paskavitz | Wayne State University School of Medicine |
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| Megan | Pino | The Ohio State University College of Medicine |
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| Yusha | Sun | University of Pennsylvania School of Medicine |
| Jonathan | Sussman | University of Pennsylvania School of Medicine |
| Maya | Talukdar | Harvard/MIT MD-PhD Program |
| Trevor | Tamura | University of Colorado MSTP |
| Hunter | Tolison | University of Colorado MSTP |
| Josephine | Trichka | Case Western Reserve University MSTP |
| Idil | Tuncali | University of Louisville School of Medicine MD/PhD Program |

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| Jude | Tunyi | The Ohio State MSTP |
| Jacqueline | Turner | University of Colorado MSTP |
| Kaylee | Tutrow | Indiana University School of Medicine |
| Amanda | Velez | University of North Carolina at Chapel Hill |
| Amitej | Venapally | Emory University School of Medicine |
| Courtney | Vetter | University of Colorado MSTP |
| Mayuri | Viswanathan | University of Chicago MSTP |
| Madison | Wahlen | University of Iowa |
| Madison | Walker | University of Colorado MSTP |
| Qi | Wang | University of Iowa Carver College of Medicine |
| Jenna | Weber | University of Utah |
| Natalie | Weed | University of Colorado MSTP |
| Margaret | Weiss | University of Illinois at Chicago |
| John | Wells | Indiana University School of Medicine MSTP |
| Clare | Wieland | University of Michigan |
| Ben | Wong | University of Colorado MSTP |
| Elizabeth | Woo | Yale School of Medicine |
| Nathaniel | Wright | University of Chicago |
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| Crystal | Young | University of Michigan MSTP |
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| Deanne | Yugawa | Yale MD-PhD Program |
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| Sherry | Zhou | Mayo Clinic M.D.-Ph.D. Program |
| Samuel | Zorn | Medical College of Wisconsin |