

SCHOOL OF MEDICINE

UNIVERSITY OF COLORADO ANSCHUTZ MEDICAL CAMPUS

HI3 Member Meeting
February 14, 2017
2:00-3:00pm
Shore Family Auditorium
Nighthorse Campbell Building

Medicine	<b>\$</b>	Q

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## ucdenver.edu/HumanImmunology



#### About

#### Faculty

HI3 Directors

HI3 Internal Advisory Committee

HI3 External Advisory Committee

HI3 Members on the Anschutz Medical Campus

Infrastructure

#### Training

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Events

#### **HIMSR**

Meet the Team

Collaborative Resources

Equipment and Services

Rates and Fee Structure

#### News

Status Reports



After decades of investigation establishing principles in animal models, it has recently become possible to treat and cure some diseases in humans by interventions that target immunological functions. Immunotherapy, described by Science magazine as the Breakthrough of the Year in 2013, has led to major changes in the standard of care for some diseases and is particularly useful in infectious disease, autoimmunity, allergy/asthma, and especially cancer.

The **mission** of the **Human Immunology and Immunotherapy Initiative (HI<sup>3</sup>)** is to develop the infrastructure to create an all-inclusive facility that provides experimental models for the preclinical testing of new candidate therapeutics, reliable immunotherapeutic production, consistent clinical trials research support, and organized immune monitoring capabilities. In addition, the HI3 will train future scientific leaders within the realm of immunotherapy and recruit exemplary faculty to the CU Anschutz medical campus that will complement and enhance existing strengths.



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## **Activities**

- Faculty Recruitment
- GMP Immunotherapeutic Production
- Training Program
- Clinical Research Program
- Translational Research Networking & Preclinical Models (TRNPM)
- Human Immune Monitoring Shared Resource (HIMSR)

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# HI3 Faculty Recruitment UPDATE

## Basic Human Immunology

- ✓ Committee formed, position posted on CU Careers
- ✓ 4 candidates will be interviewed by the end of April

# Space?

## Autoimmunity

- ✓ Committee formed, position posted on CU Careers
- ✓ HI3 Autoimmunity Program Director
- ✓ Interviews to begin in late April

#### Cancer

- ✓ Working with several cancer recruits ongoing at CU and CHCO
- ✓ Terry Fry, MD (NCI) verbal acceptance January 2017, informed NCI of upcoming departure, proposed start January 2018

## Data Scientist/Bioinformatician

- ✓ Committee formed, position posted on CU Careers
- ✓ 3 candidates will be interviewed by the end of March



## HI3 GMP Immunotherapeutic Production UPDATE

- Collaborate with GMP facilities toward the production of clinical grade biological reagents and cell-based immunotherapeutic products
  - ✓ Clinimmune
  - ✓ The Gates Center for Regenerative Medicine
- Facilitate the use of campus CLIA labs for monitoring patient responses used for clinical decision-making
  - ✓ Exsera BioLabs complement and autoimmune diagnostics
  - ✓ Colorado Molecular Correlates Laboratory (CMOCO) anatomic pathology
  - ✓ Clinimmune flow cytometry, cell sorting, histocompatibility

## HI3 Training Program UPDATE

- Develop and establish training programs across the training continuum at the pre-doctoral, post-doctoral, and junior faculty level
  - ✓ Establish fellowships to support training and research for future leaders in immunotherapy – 2 PhD candidates, 2 post-doctoral fellows, 1 junior faculty
  - ✓ Work with selected faculty and established campus services to provide educational resources
- Form a Training Program Subcommittee to play a role in the design and structure of the program \*1

## Are you interested?

Please see me - Aimee Bernard



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## HI3 Clinical Research Program UPDATE

## HI3 Clinical Research Program Director

✓ Physician scientist (MD) to develop and oversee program

## Clinical Research Program Manager \*2

- ✓ Successfully guide HI3 investigators through the complicated web (CRSC/CCTSI, CTRC, CCRO/RI, UCCC, CHCO CC) of established CU clinical research support services
- ✓ Provide expertise and coordination with campus clinical research services to HI3 research teams to effectively operationalize clinical research
- ✓ Establishes and maintains bi-directional communication and collaboration between HI3 research teams and campus clinical research services from initiation to completion of clinical studies

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## Translational Research Networking & Preclinical Models Facility UPDATE

- Provide a nexus for multiple aspects of translational immunology research
  - o Director: Roberta Pelanda, PhD
  - o Manager Humanized Mouse Core HI3: Julie Lang, PhD

## **Services include:**

- ✓ Enabling and promoting collaboration among investigators along the continuum of basic-clinical-translational research by networking and/or matchmaking clinicians, clinician/scientists, and basic scientists with shared interests
- ✓ Establishing mechanisms that ensure availability of human tissue for research
- ✓ Developing and maintaining **preclinical mouse models** for testing of candidate therapeutics (e.g. human immune system mice or **hu-mice**) \*3

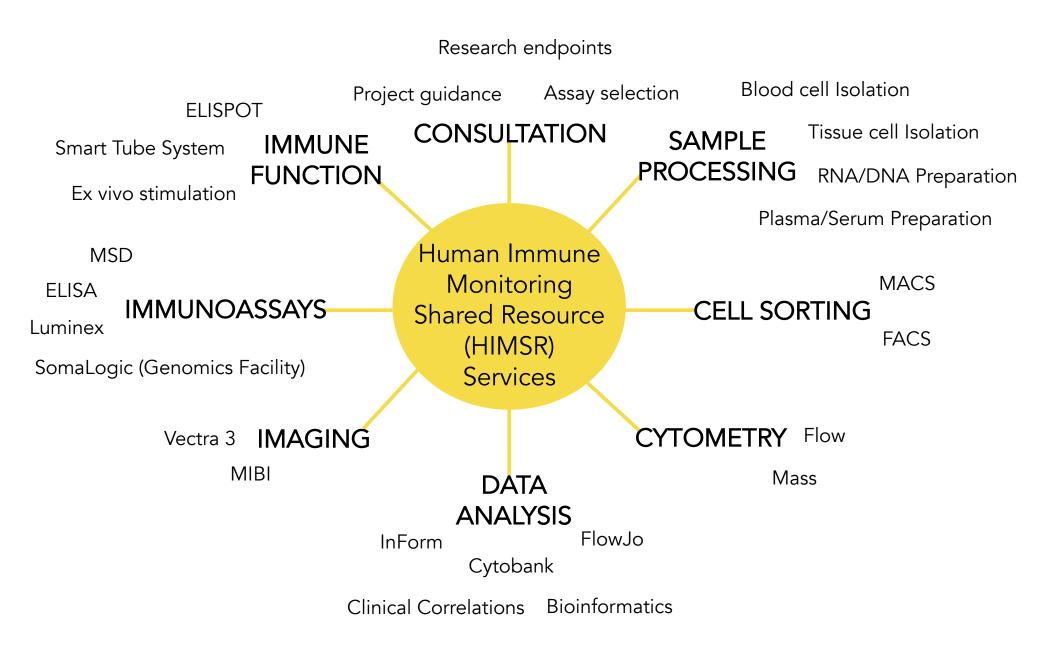


## Human Immune Monitoring Shared Resource (HIMSR) UPDATE

#### HIMSR Team Members

- o Director: Jill Slansky, PhD
- o Assistant Director: Kim Jordan, PhD
- o Experimental Design Consultant: Elena Hsieh, MD
- o PRA/Histology Specialist: TBD
- Flow cytometry and protein purification specialist: Jennifer McWilliams,
   PhD
- o Immuno&Micro Flow Facility Manager: Erin Kitten, BS

## Human Immune Monitoring Shared Resource (HIMSR) Services



### **HIMSR Consultation**

- Intentions
  - o Provide immunology expertise
  - o Discuss project goals and HIMSR assistance
  - o Identify and design feasible immune monitoring experiments
  - o Discuss funding sources and budgets
- What we've been working on
  - o Study design
  - o Experimental descriptions for methods sections
  - o Letters of support and collaboration
  - o Grant budgets
  - o Grant facilities pages

IMMUNE FUNCTION

IMMUNOASSAYS

IMMUNOASSAYS

IMAGING

CONSULTATION SAMPLE PROCESSING

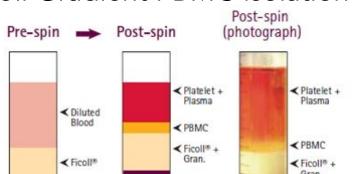
Human Immune Monitoring Shared Resource Services

CYTOMETRY DATA ANALYSIS

 Currently, we are working with 20+ interested groups at various funding stages and across different services

## HIMSR Sample Processing

Ficoll Gradient PBMC isolation



CPT tube PBMC isolation



Plasma and PBMC aliquots



Tissue cell isolation- Miltenyi gentleMACs

< RBC



RNA and DNA isolation



Temporary cryopreservation, immune monitoring assays, genomics, proteomics

### **HIMSR Immune Function**

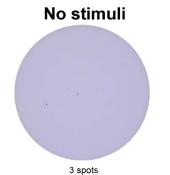
#### Ex vivo Stimulation: SMART Tube

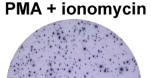


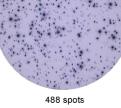
- Whole blood or PBMC stimulation
- Preservative added after incubation, ready for freezing.
- Study samples thawed together and stained (surface markers, signaling molecules, or cytokines)

#### In Development

#### **ELISPOT**

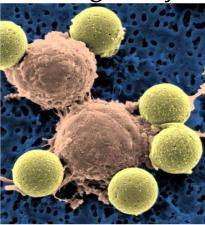






**Proliferation** 

## Killing Assays



- Flow-based
- Incucyte
   (UCCC Tissue
   Culture core)

#### Flow cytometry

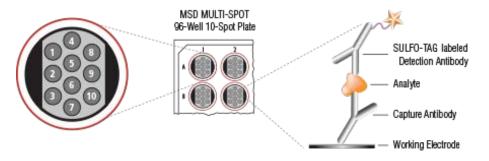


Mass cytometry



## HIMSR Immunoassays





- Parsec automated instrument
- Up to 10 analytes per 25 ul of plasma
- ~40-plex validated assays
- \$75,000 in free reagents once the Parsec arrives and we have space for it! \*4





- Manual preparation of plates to read at the Cancer Center Flow Core
- Up to 500 analytes per sample

# Somalogic

- Upstream sample preparation for SOMAscan assays performed in the Cancer Center Genomics Facility
- Up to 1,300 analytes per sample

## HIMSR Flow and Mass Cytometry

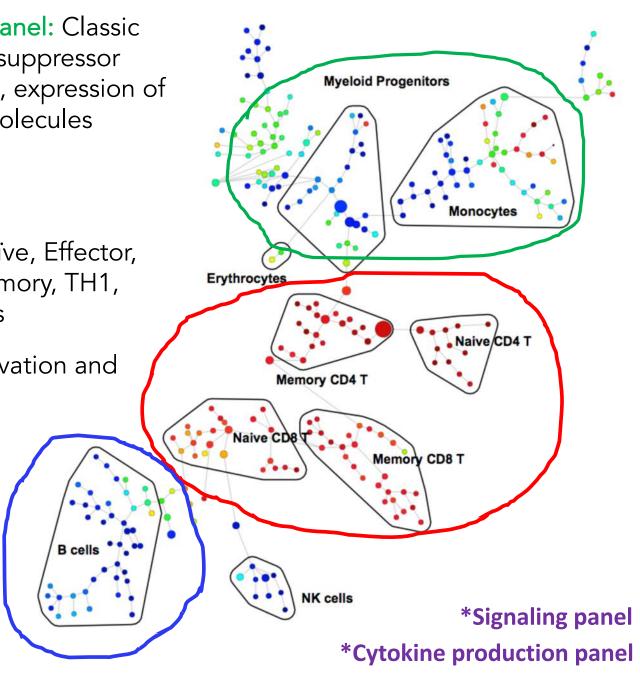
Monocyte/DC Phenotyping panel: Classic and activated monocytes, suppressor cells, dendritic cell subsets, expression of activation and inhibitory molecules

T cell Phenotyping panel: Naïve, Effector, Effector Memory, Central Memory, TH1, TH2, TH17, Regulatory T cells

T cell activation markers: Activation and

Inhibitory receptors

B cell Phenotyping panel: Immature, naïve, memory B cells, plasmablasts, isotype, activation and inhibitory molecules



## HIMSR Flow and Mass Cytometry

## T cell Phenotyping panel B cell Phenotyping panel Monocyte/DC Phenotyping panel

CD3, CD4, CD8, FOXP3, CD25, CD45RA, CD45RO, CD62L, CCR6, CXCR3, CXCR4, CCR7, CD66 IgM, IgG, IgD, IgA, CD19, CD20, CD10, CD27, CD138, CD38, CD81, CD5, CD22, CD66 Lineage, HLA-DR, CD14, CD16, CD15, CD33, CD66, CD11b, CD11c, CD1c, CD141, CD123, CD80, PDL1

#### T cell activation markers

CD3, CD4, CD8, CD66, CD45RO, Tim-3, CTLA4, CD25, LAG-3, PD-1

#### Signaling panel

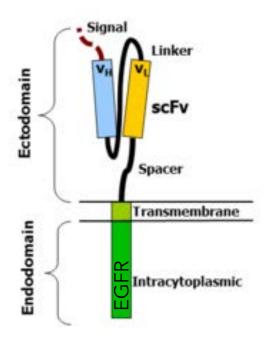
CD3, CD4, CD8, CD19, CD56, CD11b/CD66, phospho-S6, IkBa, phospho-AKT

#### Cytokine Production panel

CD3, CD4, CD8, CD19, CD56, CD11b/CD66, TNFa, IFNg, IL-4, IL-6, Granzyme B

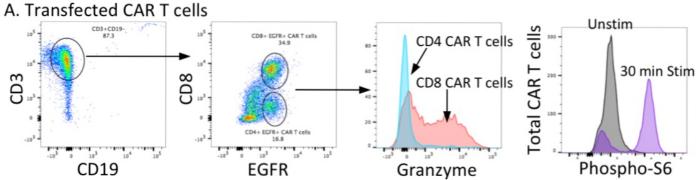
- Are you interested in adding your favorite cells markers? Contact us! \*5
- We will also provide these antibody clone names and staining protocols for investigators wishing to perform the staining in their own laboratories.

## Monitoring CAR-T cell frequency and function in ALL patients

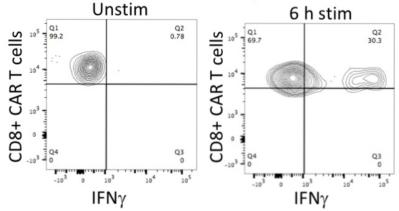


#### HIMSR will be:

- Processing peripheral blood at various time points post-infusion
- Stimulating cells (SMART tubes) and collecting plasma
- Analyzing frequency, subsets, and effector function of CAR T cells
- Measuring inflammatory cytokines in plasma



B. Healthy Donor Whole Blood spiked with CAR T cells

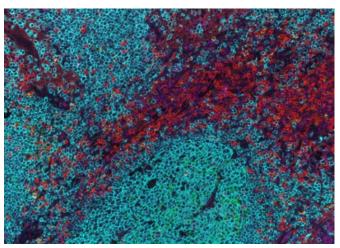


Cancer Center Immunotherapy RFA, Enkhee Purev, M.D., PI

## HIMSR Imaging - in situ visualization: Vectra 3.0 (Perkin Elmer)

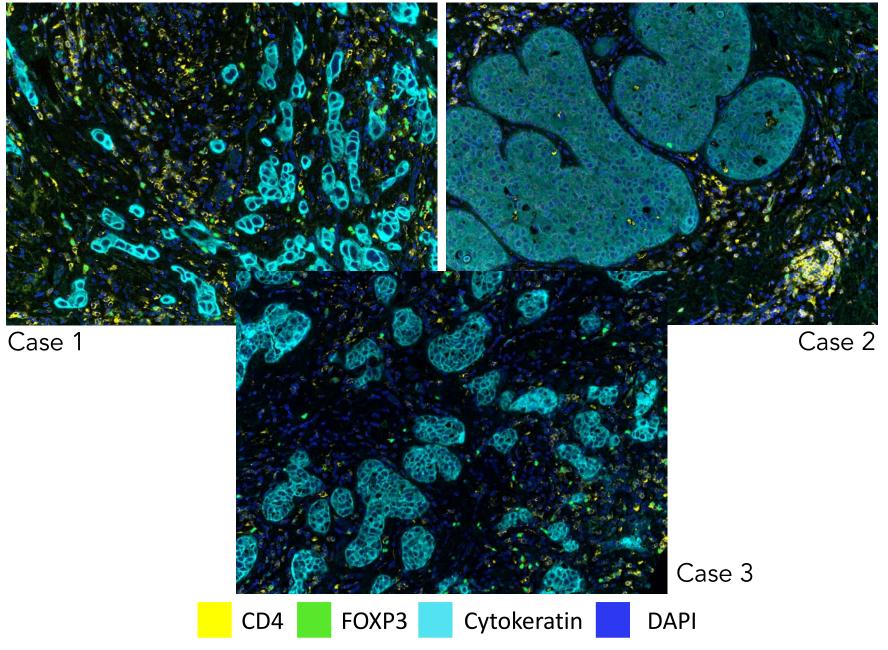
# Automated brightfield and fluorescence imaging platform





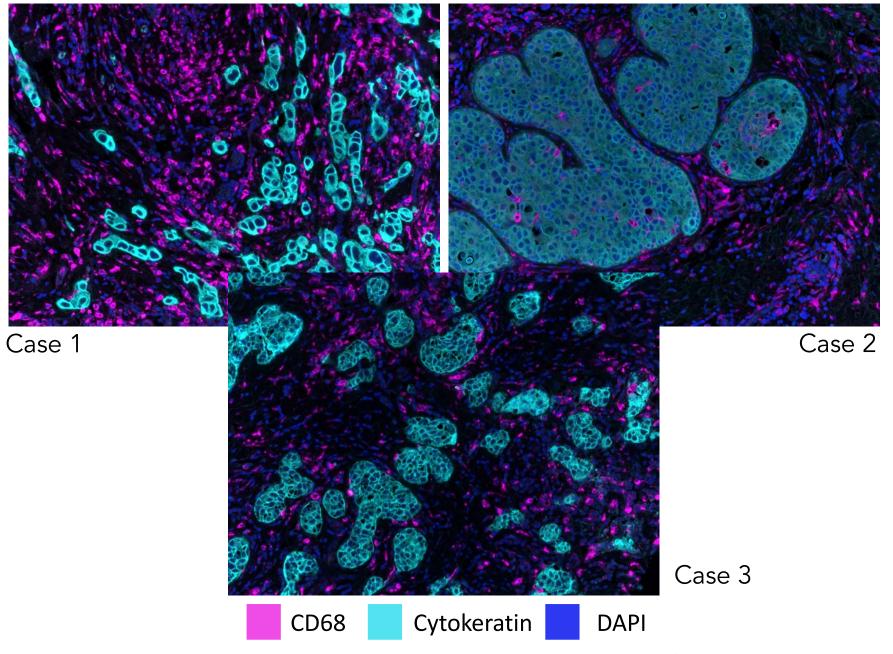
- Serial staining protocol allows for DAPI plus
   6 fluorescent tissue or cell markers
- Brightfield images (brown, red, green, H&E)
- Intended for formalin—fixed paraffinimbedded tissue (paraffin blocks)
- Whole slide scans (10x) with selected regions of interest for analyses (20x)
- Tissue Microarrays
- IRB exempt protocol submitted to obtain human tissue for staining protocol optimization (UCCC Tissue Biobanking & Processing Shared Resource)
- Working on a series of immune cell panels...
   coming soon!

Imaging – Quantification of immune infiltrate in Young Women's Breast Cancer



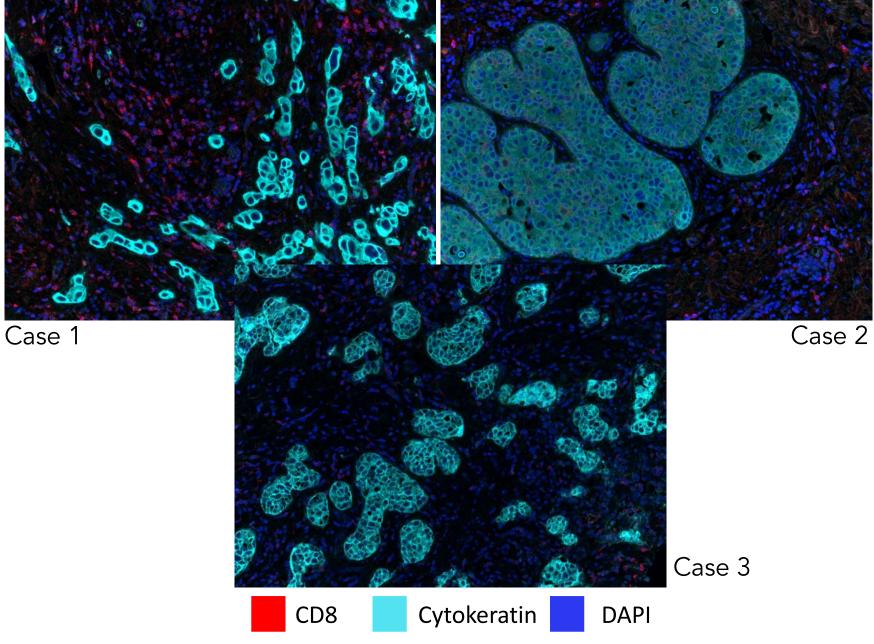
Young Women's Breast Cancer Translational Program, Dr. Virginia Borges

Imaging – Quantification of immune infiltrate in Young Women's Breast Cancer



Young Women's Breast Cancer Translational Program, Dr. Virginia Borges

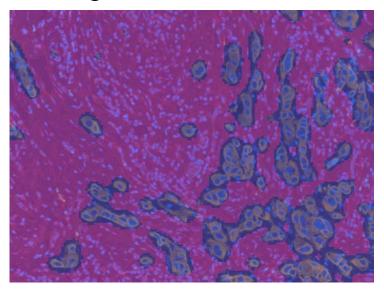
Imaging – Quantification of immune infiltrate in Young Women's Breast Cancer



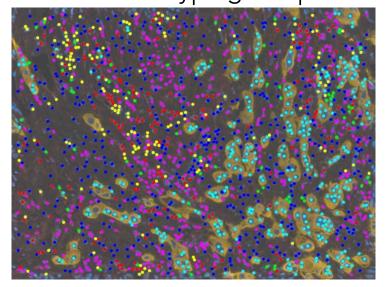
Young Women's Breast Cancer Translational Program, Dr. Virginia Borges

## Features of inForm software analyses

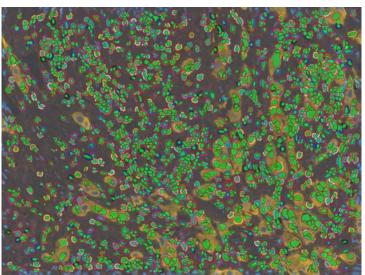
Tissue Segmentation: tumor v stroma



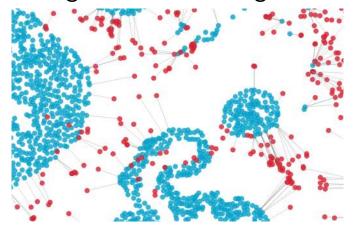
Cellular Phenotyping: frequencies



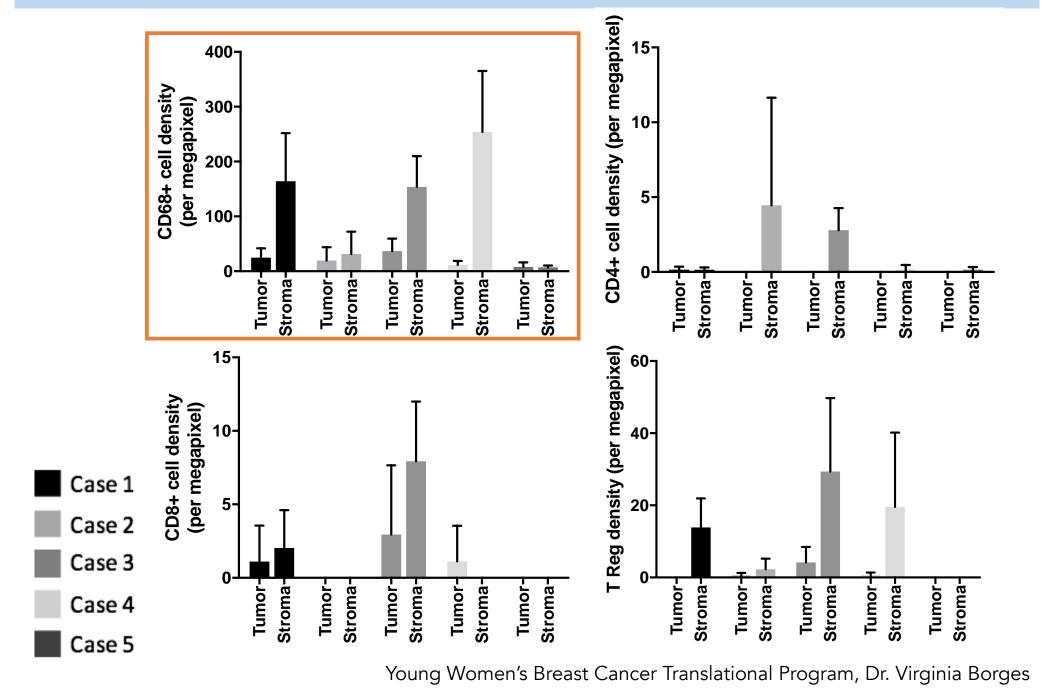
Cell Segmentation: staining intensities



- Traditional scoring function with user-set thresholds
- Spatial information (x, y coordinates)
   allowing for nearest-neighbor analysis



## Imaging and Data Analysis – Quantification of immune infiltrate in Young Women's Breast Cancer



# Example HIMSR project Immunomonitoring of metastatic colorectal cancer patients treated with anti-PD-1 therapy

1. Ficoll Gradient PBMC isolation

Pre-spin Post-spin (photograph)

Platelet + Plasma

Platelet + Plasma

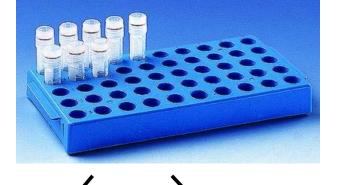
Platelet + Plasma

PBMC

Ficoll® + Gran.

RBC

2. Temporary storage of plasma and PBMC



5. *In situ* immune infiltrate in biopsies



3. Flow cytometry



4. Cytokine Array



## HIMSR Data Analyses and Bioinformatics

## Self-use analysis stations:

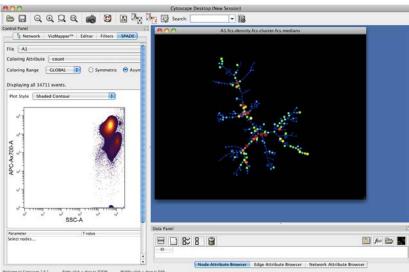
1. Image analysis



P18-8402a (Slansky lab)

### 2. Flow and Mass Cytometry analysis

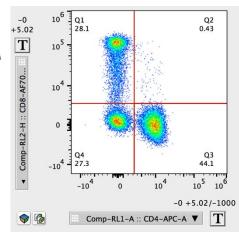






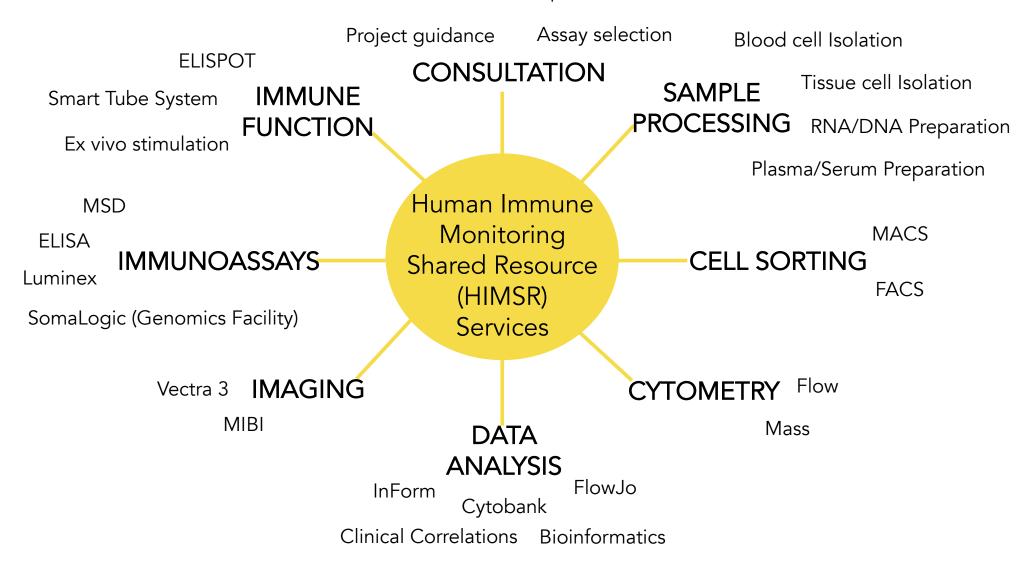
## Data analysis services:

- Flow: basic marker analysis and expression level
- Mass Cytometry: data normalization, barcode unmixing, marker analysis and expression level
- Vectra: images only, tissue and cell segmentation, phenotyping, scoring
- Depth of analyses dependent on investigator's need
- Bioinformatician



## Human Immune Monitoring Shared Resource (HIMSR) Services

Research endpoints



# Interested in working with HIMSR? Contact Kim Jordan and/or visit iLabs